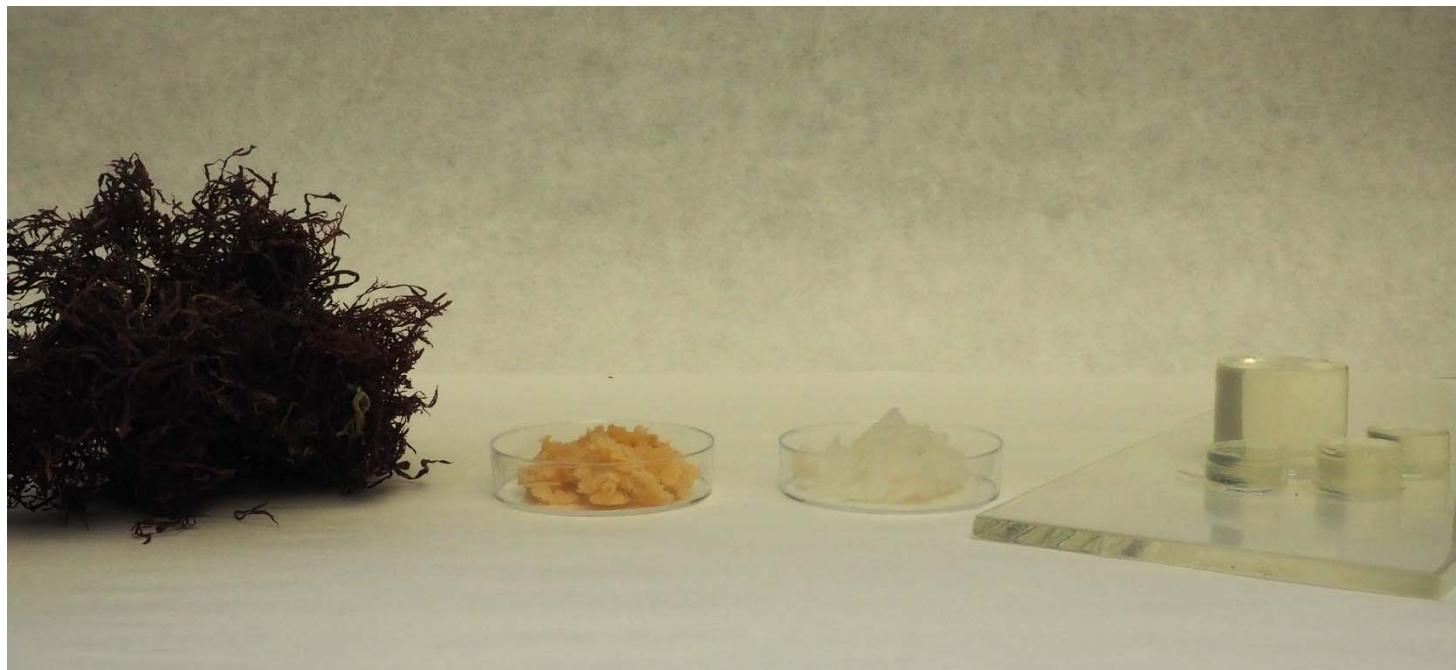
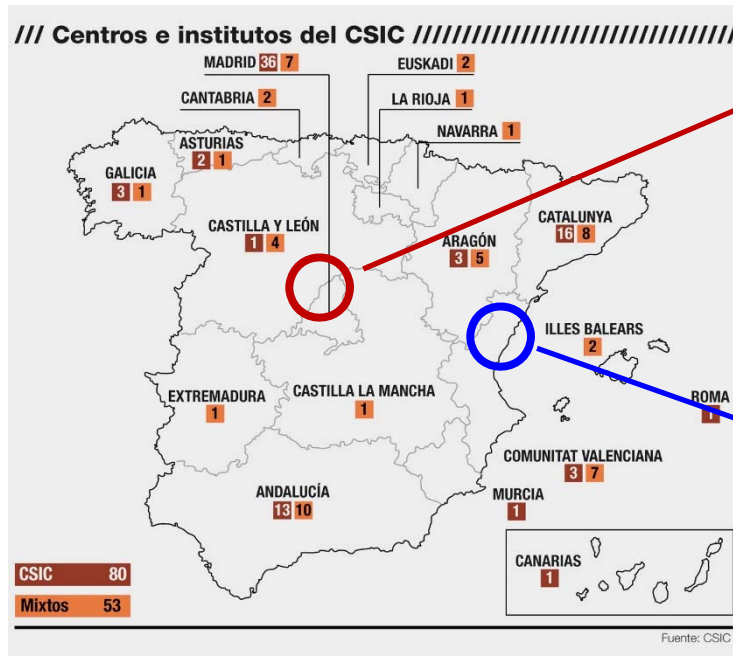


# The potential of small angle scattering techniques to study the structure of polysaccharide gels



## Spanish Council for Scientific Research (CSIC)



### Institute of Food Science Research (CIAL)

- Impact of food structure and polysaccharide-protein interactions on gastrointestinal digestion
- Development of polysaccharide-protein products resistant to gastric digestion

### Institute of Agrochemistry and Food Technology (IATA)

- Extraction of polysaccharides from marine biomass
- Development of polysaccharide-based gel-like structures with interest for food industry



University of Queensland  
(UQ)

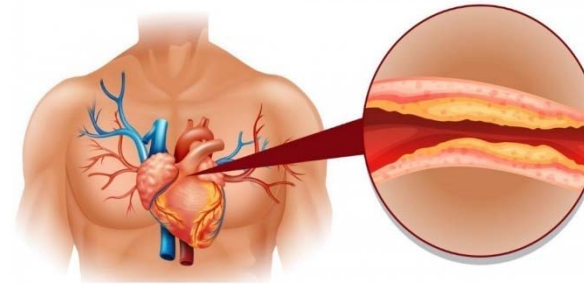
Plant Cell Walls  
ARC Centre of Excellence



Australian Nuclear Science and Technology Organisation  
(ANSTO)

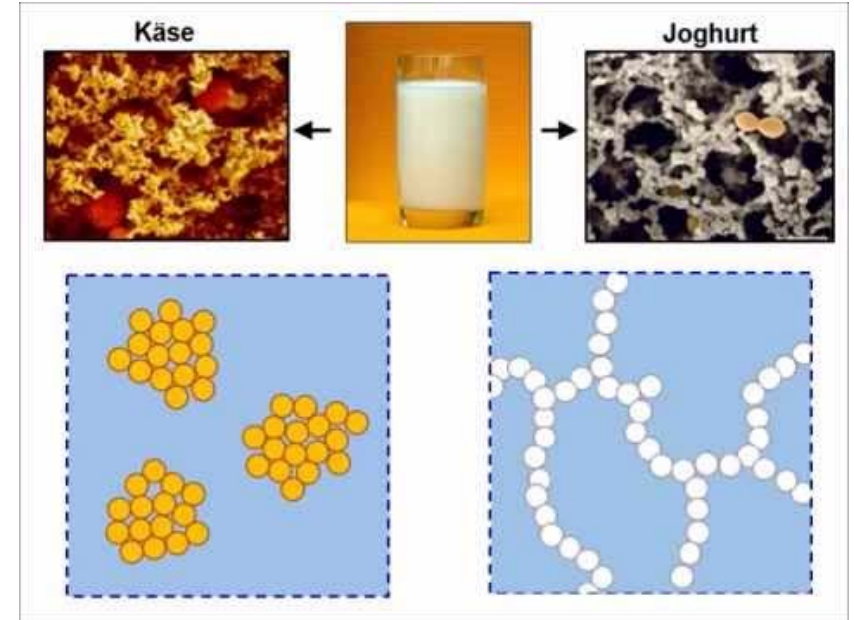
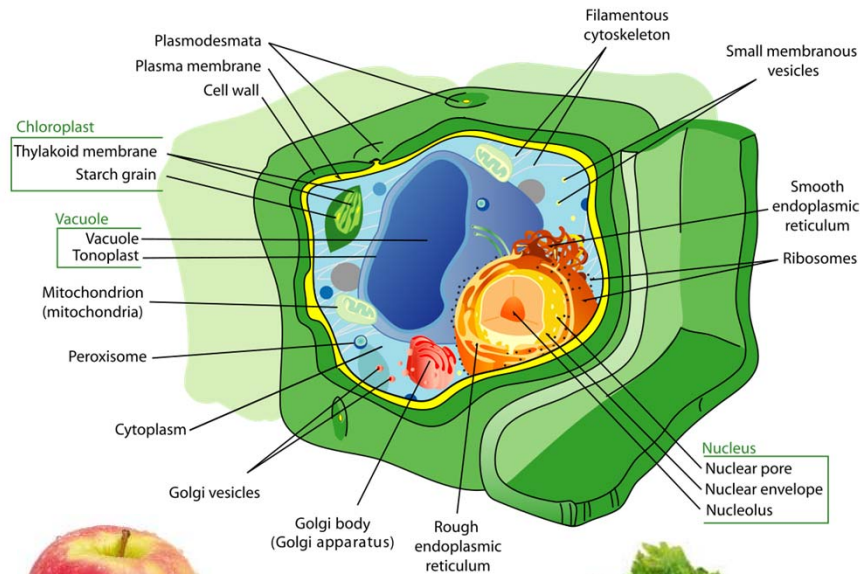


# Scattering techniques in Food Science



- Demands on the food market push for the development of **new products**: functional foods, minimally processed foods, dietetic foods, products for specific dietary requirements...
- Consumers are more concerned about the relationship between **diet and health**
- Knowledge of **food structure** is essential from an industrial (design of new products) and scientific (nutrition and health impact) perspective

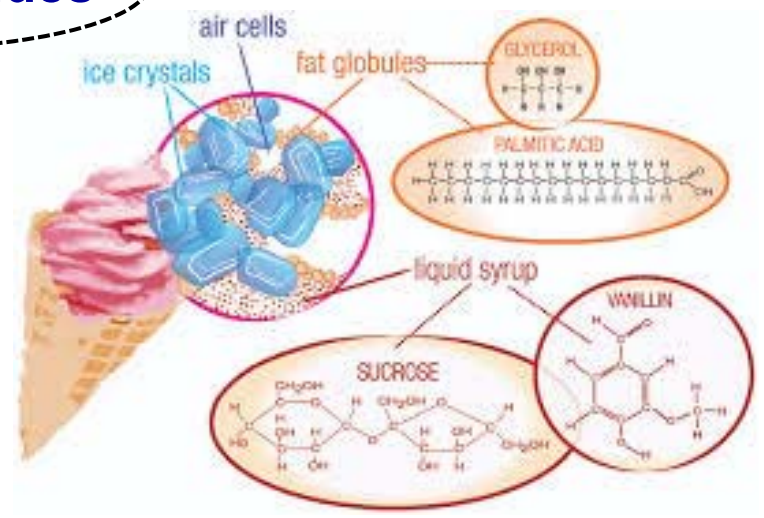
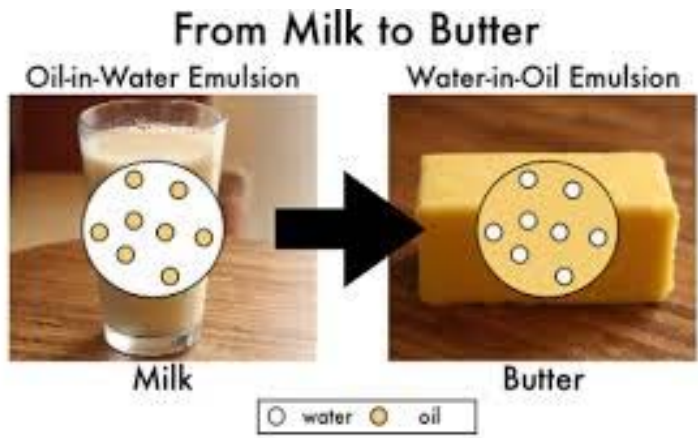
# Scattering techniques in Food Science



**Polysaccharides**

**Proteins**

**Fats**



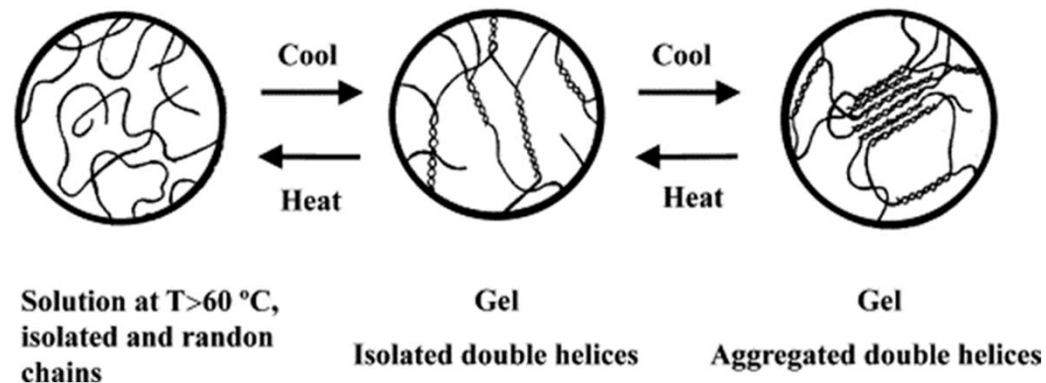
# Scattering techniques in Food Science

## Polysaccharides

- Major components in **fruits and vegetables**
- **Food ingredients:** thickeners, gelling agents, encapsulants, additives for low calorie products, etc

Source	Examples
<i>Higher plants</i>	Cellulose, hemicelluloses, starches, pectins
<i>Seaweed</i>	Agars, alginates, carragenaans
<i>Microorganisms</i>	Xanthans, pullulan, gellan, curdlan

- Many of them are found in nature in **hydrated systems**
- Might present **crystalline** (impermeable to water) or **amorphous** (water-soluble) structures

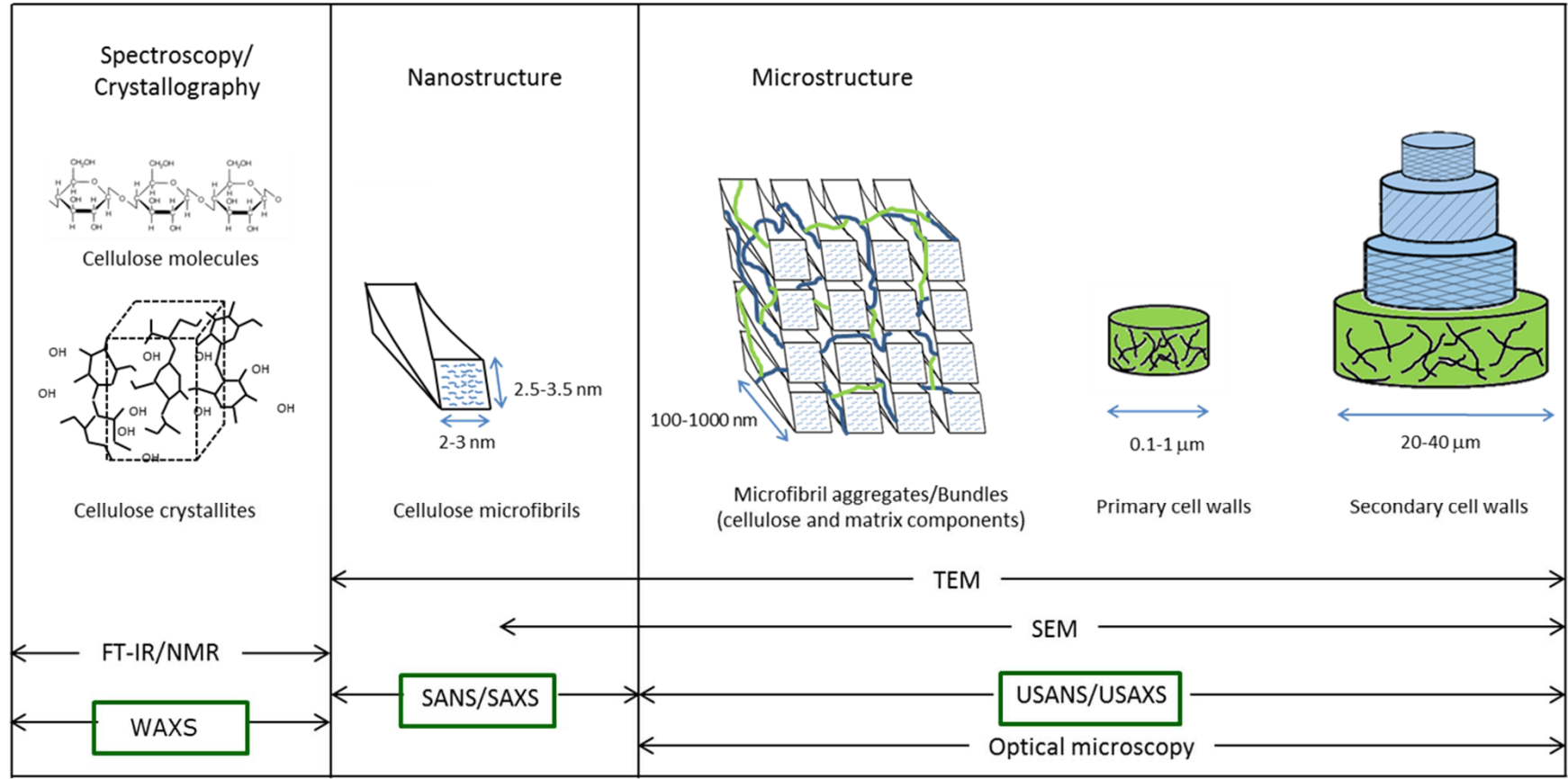


# Scattering techniques in Food Science



**Cellulose**

10<sup>-11</sup> m      10<sup>-9</sup> m      10<sup>-7</sup> m      10<sup>-5</sup> m      10<sup>-3</sup> m      10<sup>-4</sup> m

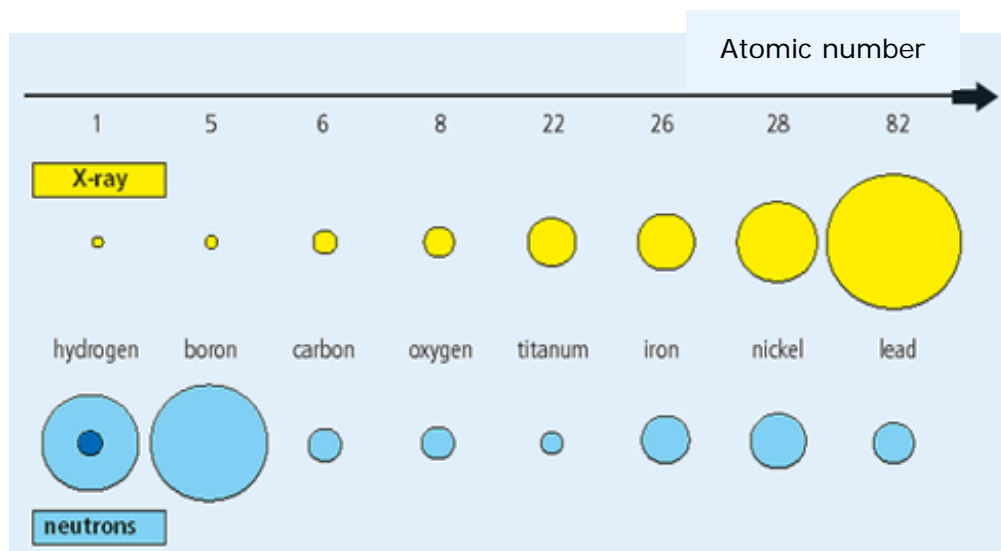


Martínez-Sanz, M., Gidley, M. J., & Gilbert, E. P. *Carbohydrate polymers* (2015)

# Scattering techniques in Food Science

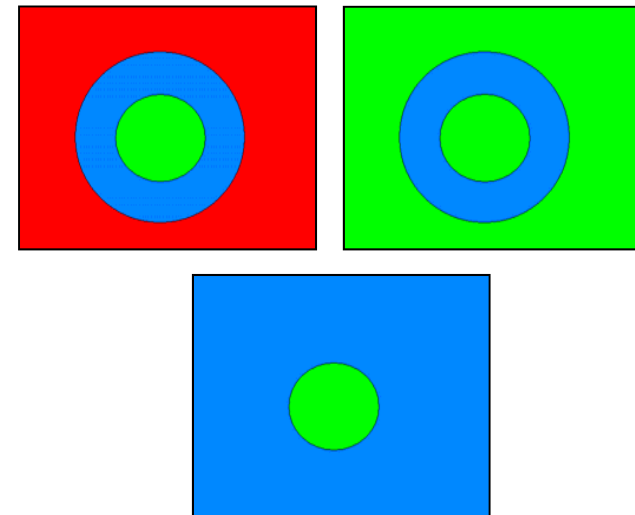
## X-Rays

- Scattered by the electrons present in an atom  
→ Sensitive to variations in electron density
- Scattering intensity increases with atomic number
- Hydrogens are almost “invisible”



## Neutrons

- Scattered by the atomic nuclei
- Different scattering intensity for isotopes of the same element (H vs D)
- Very sensitive to hydrogens
- Phase differentiation by “contrast variation”



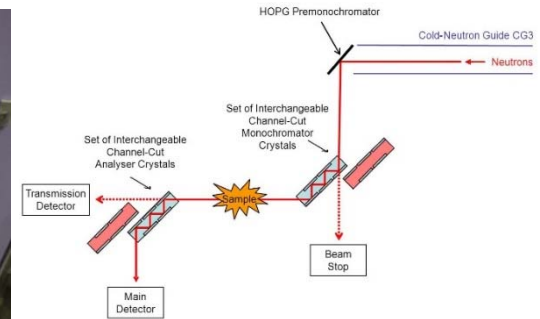
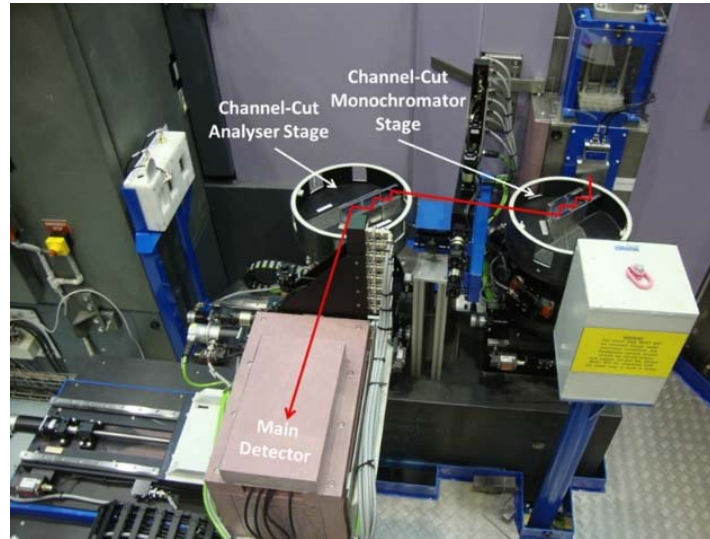
*Source of radiation selected on the basis of sample composition and structural features to be probed*

# Scattering techniques in Food Science

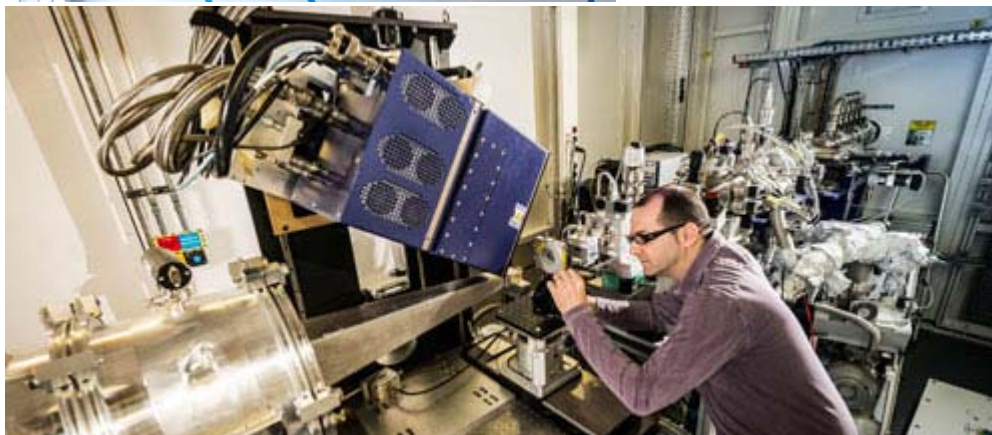
## SANS



## USANS



**Ansto**  
Nuclear-based science benefiting all Australians



**ALBA**  
Synchrotron  
Simultaneous SAXS/WAXS

**BL11 - NCD-SWEET**

# AGAR-BASED HYDROGELS

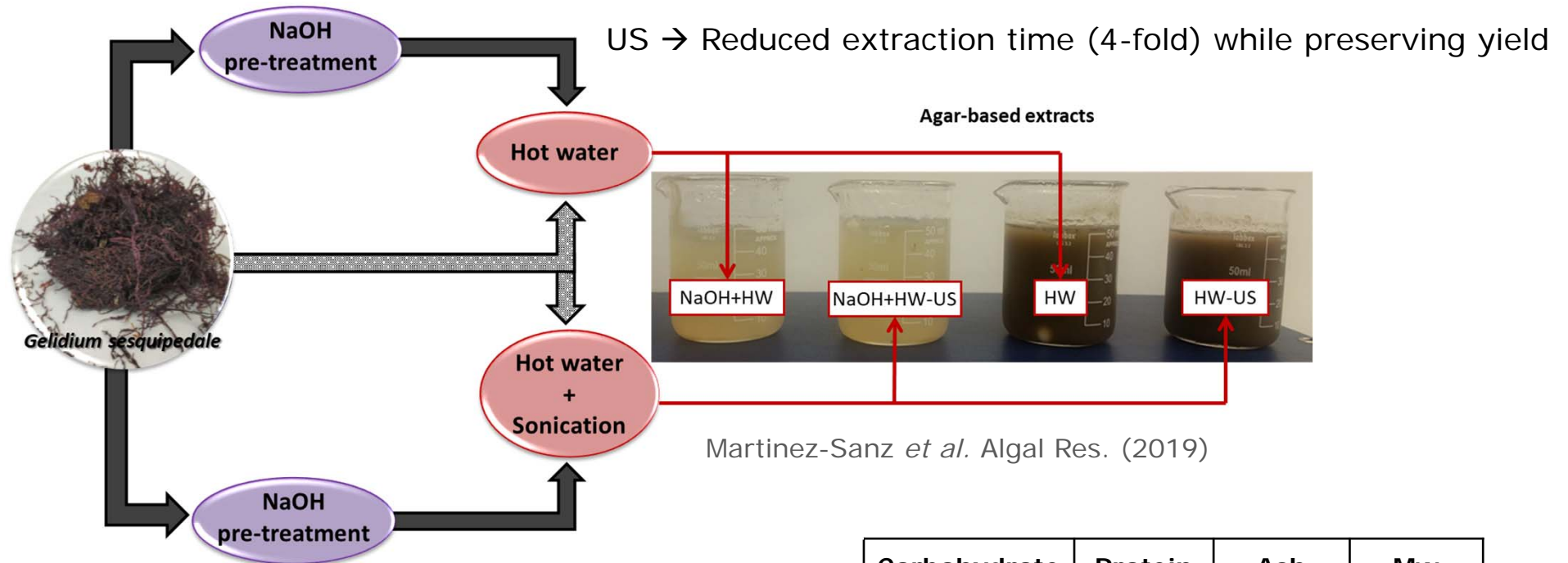


Martínez-Sanz et al. *Carb. Polym.*(2020)



# Agar-based hydrogels

Production of **agar-based extracts** from *Gelidium* using simplified extraction protocols



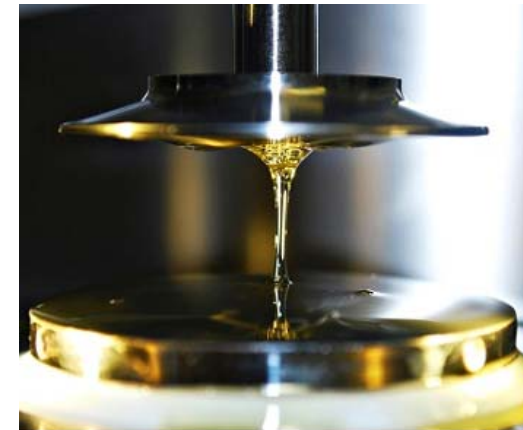
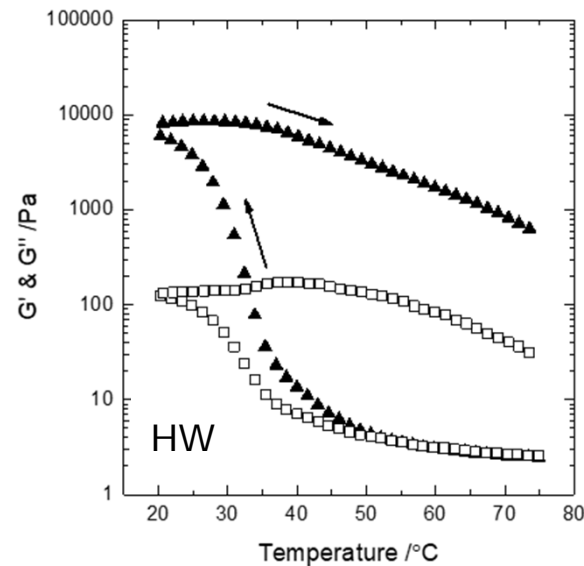
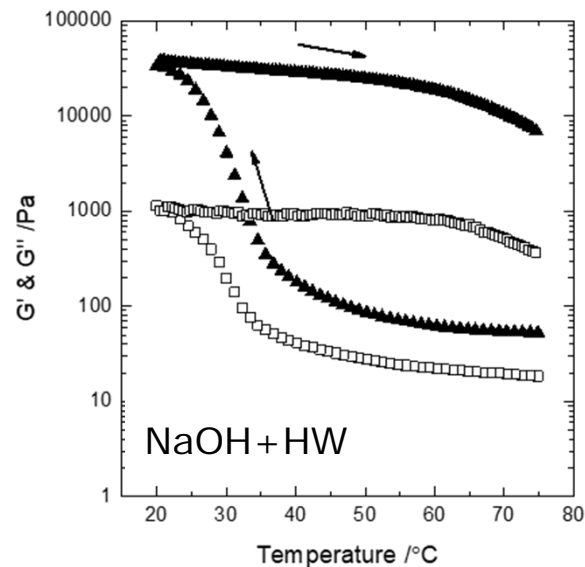
- Pre-treatment produced more purified agars, but led to partial degradation (lower  $M_w$ )
- **Non-purified extracts** are richer in proteins, minerals and polyphenols → ↑↑ **Bioactivity**

	Carbohydrate (%)	Protein (%)	Ash (%)	Mw (kDa)
<b>Commercial</b>	83	1.5	4.3	196
<b>HW</b>	42	<b>10.6</b>	<b>34.7</b>	840
<b>HW-US</b>	30	<b>11.3</b>	<b>36.4</b>	485
<b>NaOH+HW</b>	66	4.0	11.9	487
<b>NaOH+HW-US</b>	51	7.3	13.9	265

# Agar-based hydrogels



- Gelation mediated through the formation of agarose **double helices** and **bundles** upon cooling

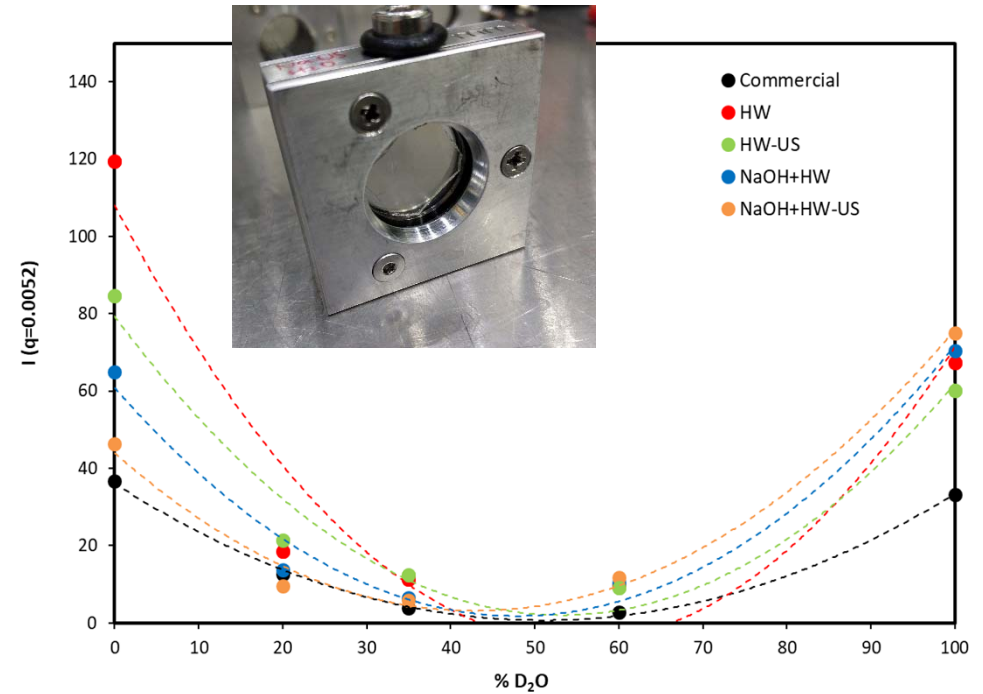
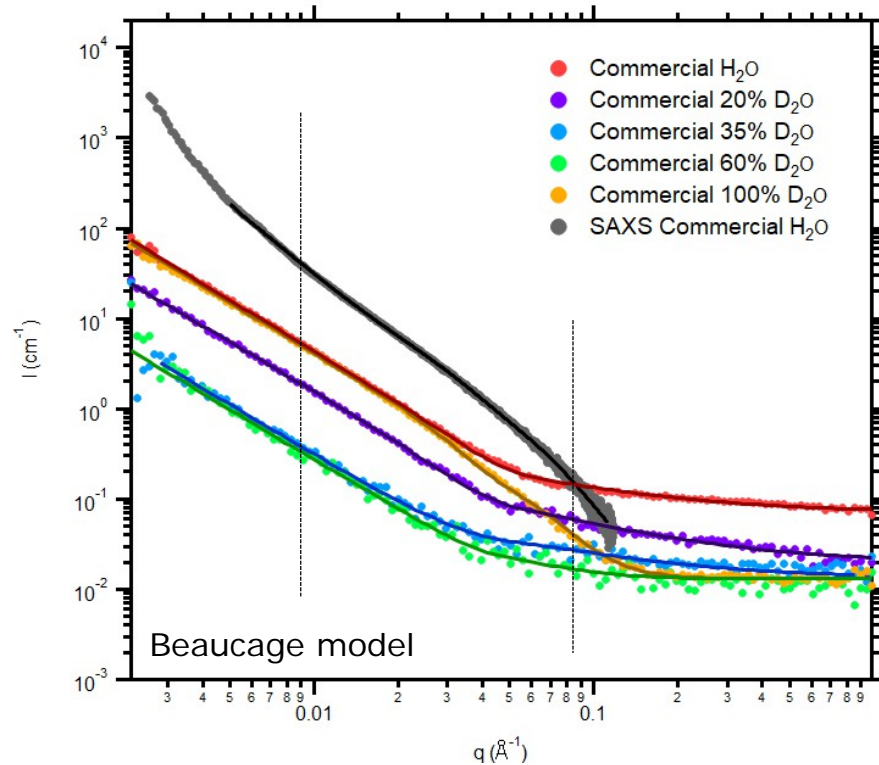


- **Similar gelling T** and mobility of cross-linking points
- Non-purified extracts form **softer gels**

- No melting upon heating
- Large syneresis

# Agar-based hydrogels

SAXS + SANS contrast variation of gels



Greater deviation from the theoretical behaviour in the non-purified agars due to (1) OH/OD exchange and (2) impurities

## Aggregation of agarose double helices:

$R_g = 5-6$  nm for the non-purified agars

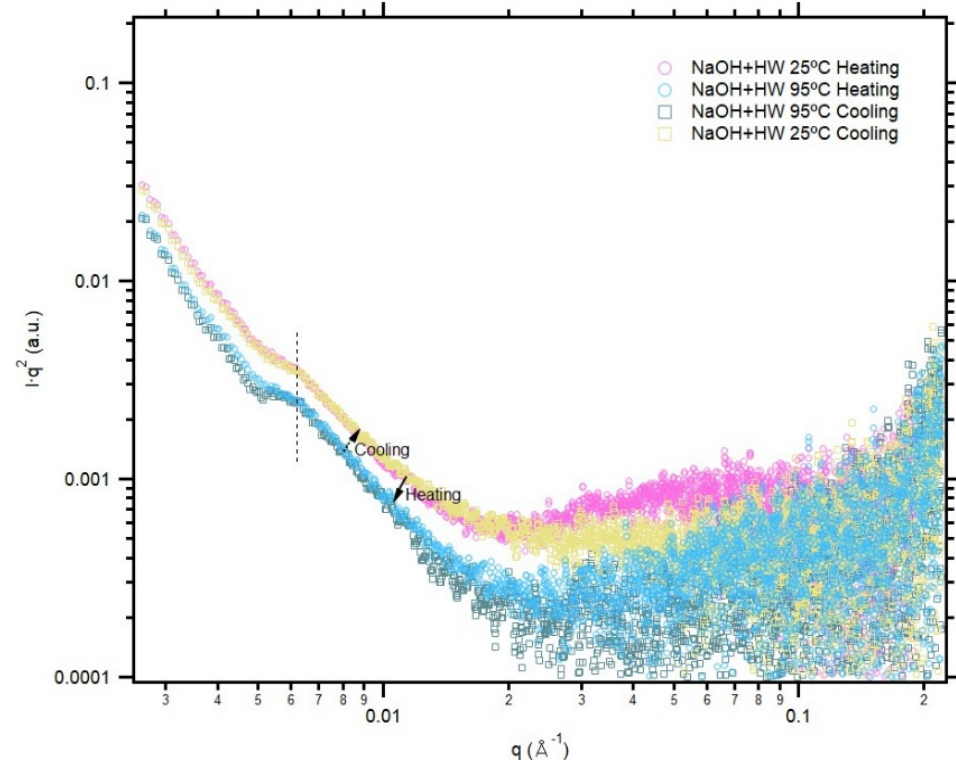
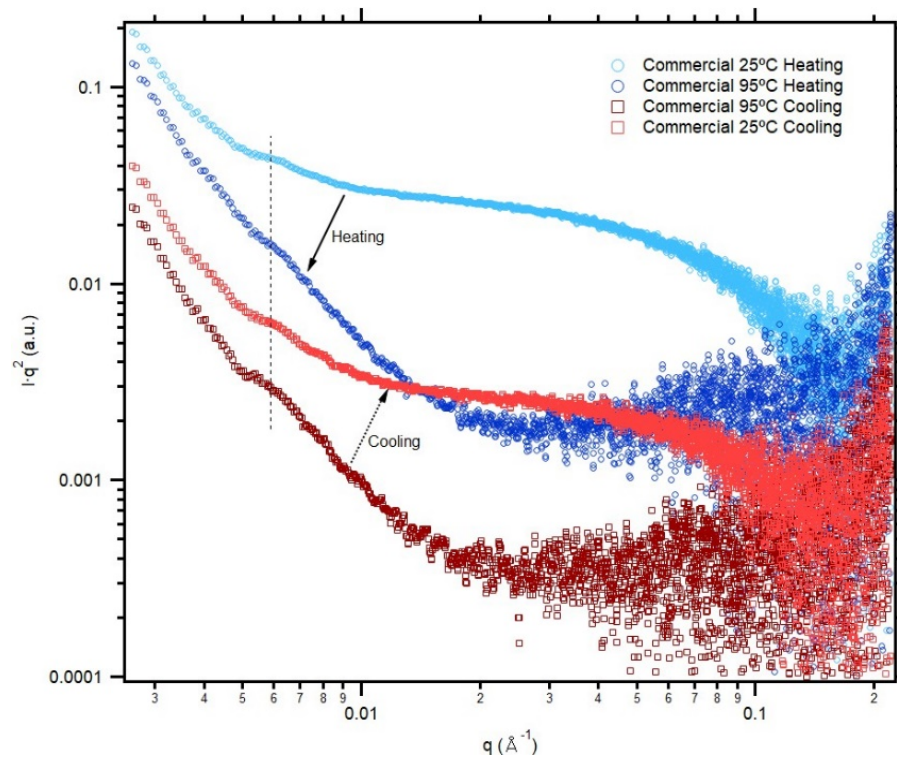
$R_g = 7-10$  nm for the purified agars

$$\text{SLD Agarose} = 2.16 \cdot 10^{10} \text{ cm}^{-2}$$

	Commercial	HW	HW-US	NaOH+HW	NaOH+HW-US
Contrast match (% D <sub>2</sub> O)	51.0	54.9	53.2	47.8	42.8
SLD (10 <sup>10</sup> cm <sup>-2</sup> )	2.96	3.23	3.11	2.74	2.39

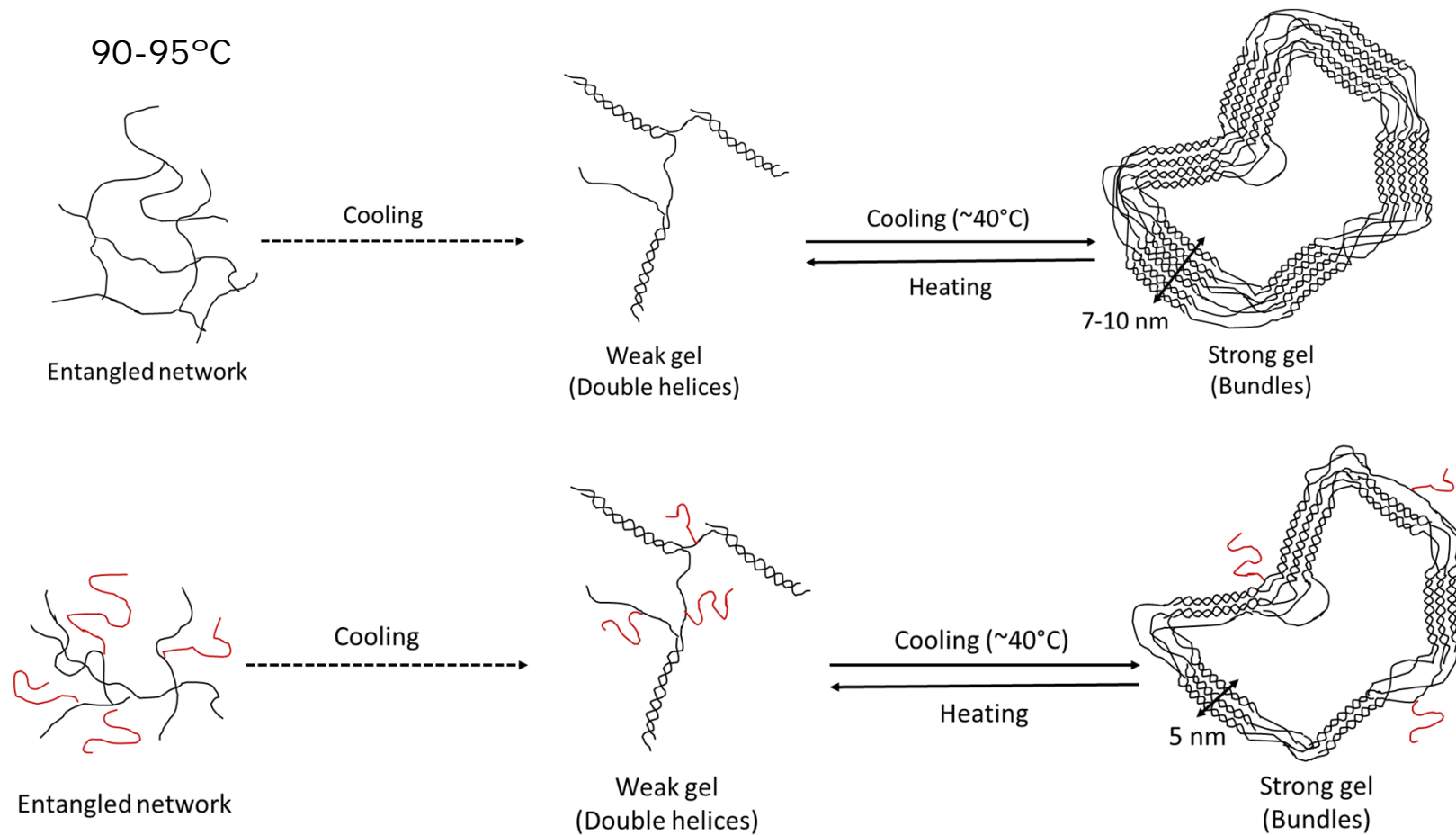
# Agar-based hydrogels

## SAXS Temperature-resolved experiments



- No real melting transition but **disruption** of agarose **double-helix aggregates**.
- Aggregates are stable even at 95°C in the more purified samples.
- The disruption of aggregates is **not completely reversible**.

# Agar-based hydrogels



- ↑Agar content → Greater extent of chain association (thicker agarose bundles) → Stronger and stiffer hydrogels
- Impurities did not impair the interconnectivity in the hydrogel network

# CARRAGEENAN HYDROGELS

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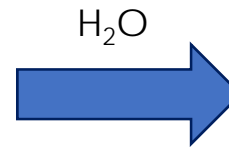
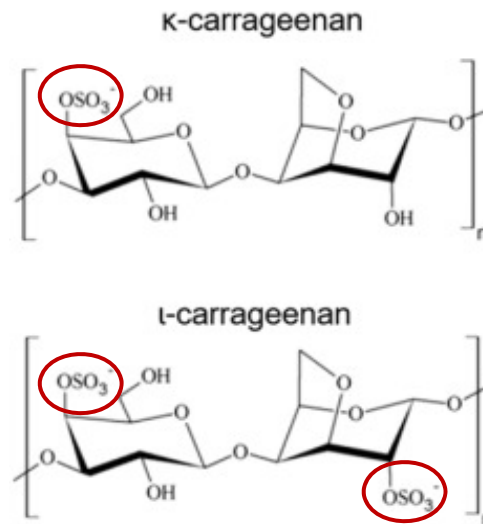
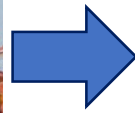


Fontes-Candia et al. *Algal Res.*(2020)

# Carrageenan hydrogels



Red seaweed



H<sub>2</sub>O

KCl  
CaCl<sub>2</sub>

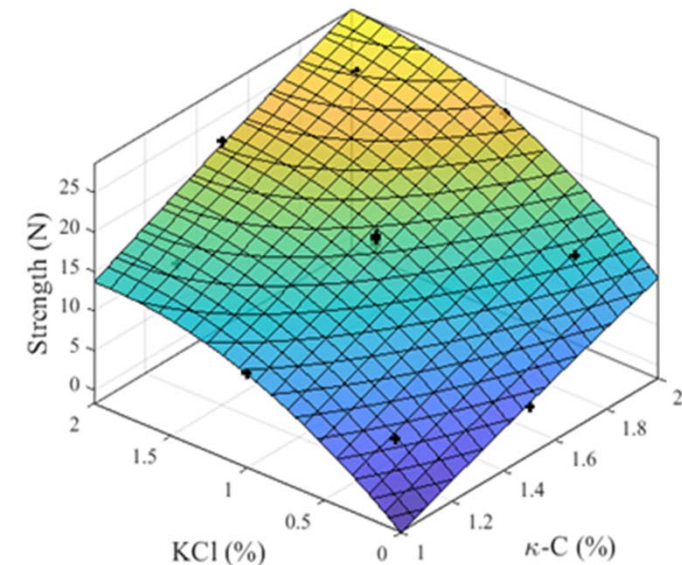


Gel strength measurements

- **Carrageenan** and **salt concentration** have a strong impact on hydrogel strength
- ↑ **sulphate content** in λ-C → **weaker hydrogels**
- K<sup>+</sup> had a greater impact in κ-C, whereas Ca<sup>2+</sup> worked better for λ-C

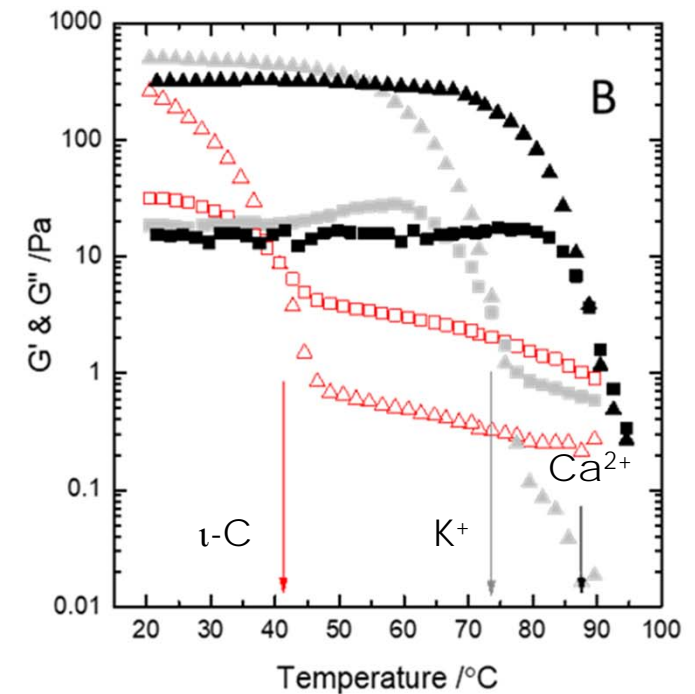
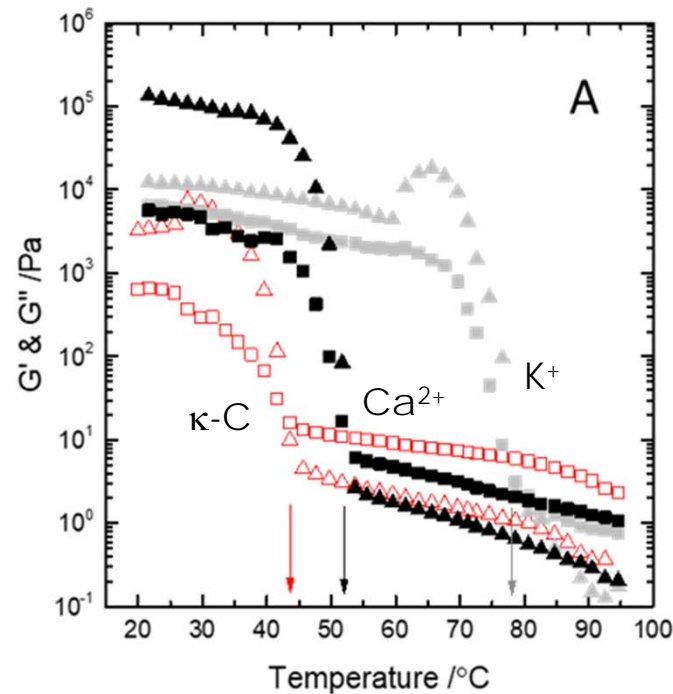
2% κ-C + 2% KCl → 29 N

2% λ-C + 1% CaCl<sub>2</sub> → 0.4 N



# Carrageenan hydrogels

2% carrageenan + 1.5% salt



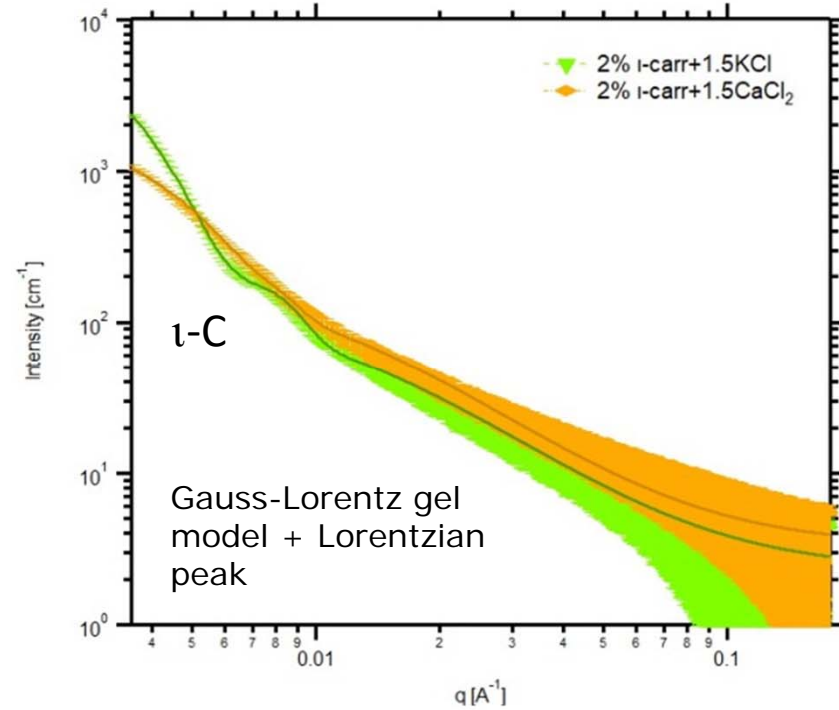
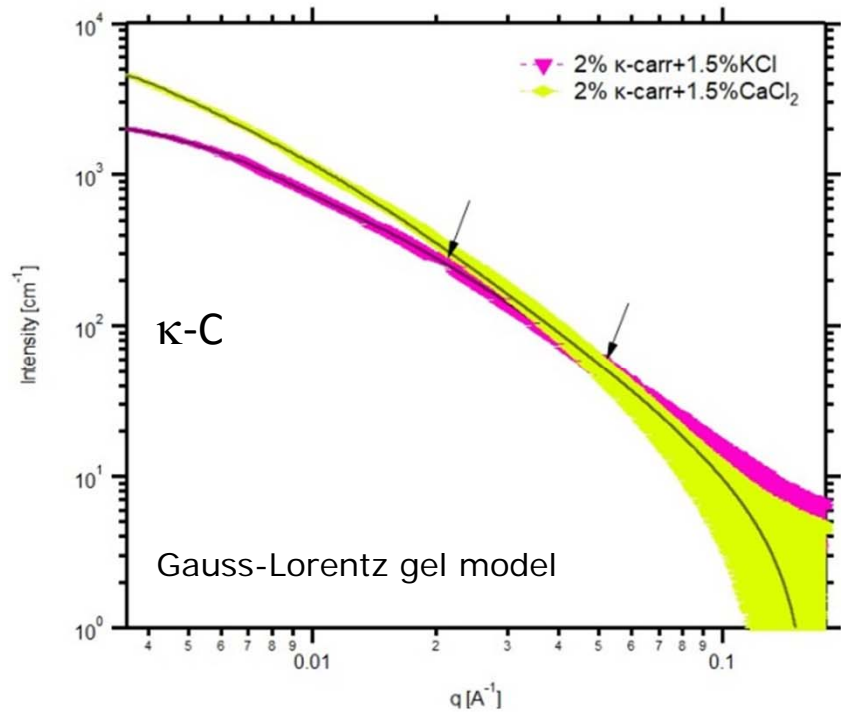
➤ Addition of salts → Promoted association of carrageenan chains → Higher gelation T

→ Greater hydrogel strength

➤  $\kappa$ -C Tgel:  $\kappa\text{-C} < \text{Ca}^{2+} < \text{K}^+$

➤  $\iota$ -C Tgel:  $\iota\text{-C} < \text{K}^+ < \text{Ca}^{2+}$

# Carrageenan hydrogels

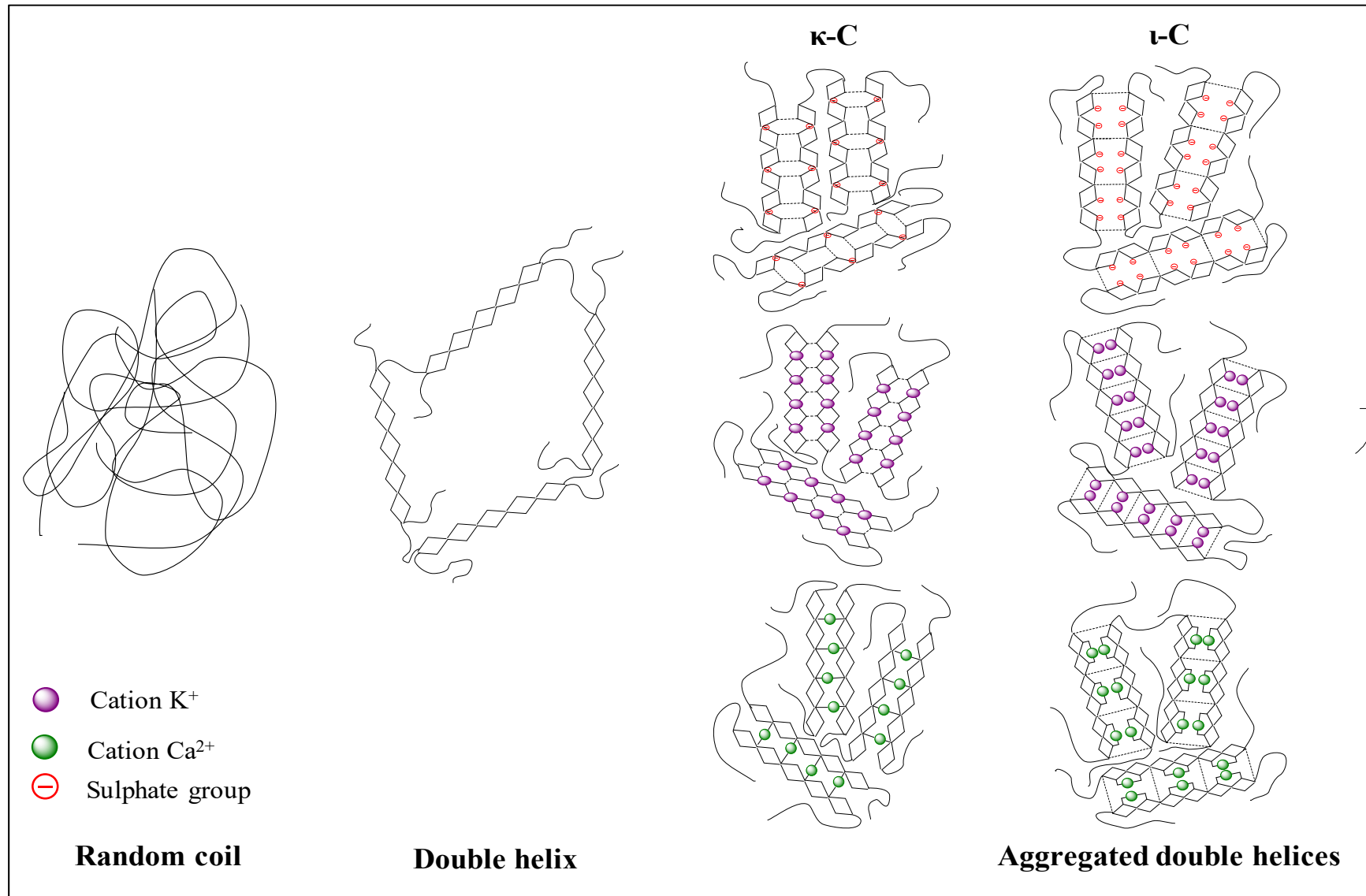


mesh size	2% $\kappa$ -C + 1.5% KCl	2% $\kappa$ -C + 1.5% $\text{CaCl}_2$	2% $\iota$ -C + 1.5% KCl	2% $\iota$ -C + 1.5% $\text{CaCl}_2$
$l_G$	1236	3091	11123	1910
$\Xi$ (nm)	25.1	41	50.4	35.14
$l_L$	1314	4888	113	172
$\xi$ (nm)	9.5	17.7	8.5	9.43
$l_0$	---	---	77	19
$d$ (nm)	---	---	85.4	77.4

double helix size

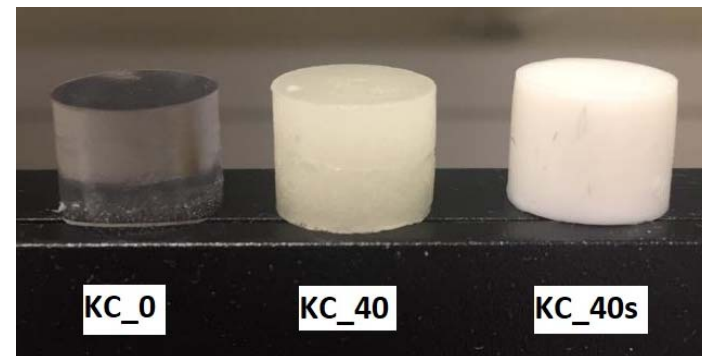
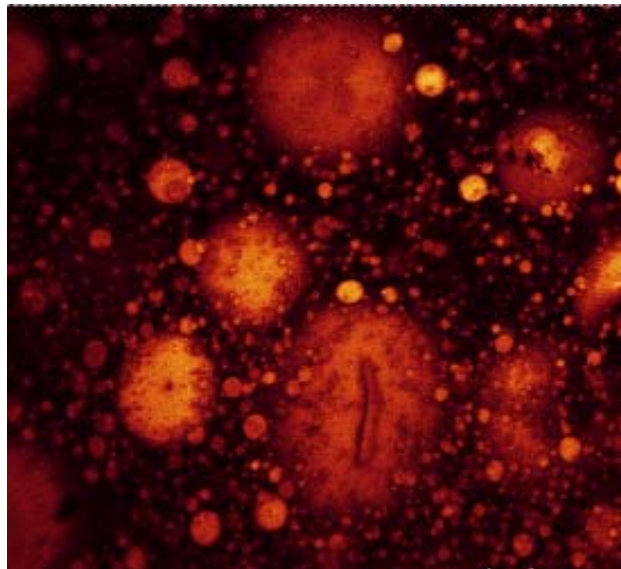
- More compact and ordered  $\kappa$ -C structures with  $\text{K}^+$
- More packed  $\iota$ -C structures with  $\text{Ca}^{2+}$
- The nature of the bridges holding together the bundles is different depending on the carrageenan and the valency of the salt

# Carrageenan hydrogels



# POLYSACCHARIDE-BASED EMULSION GELS

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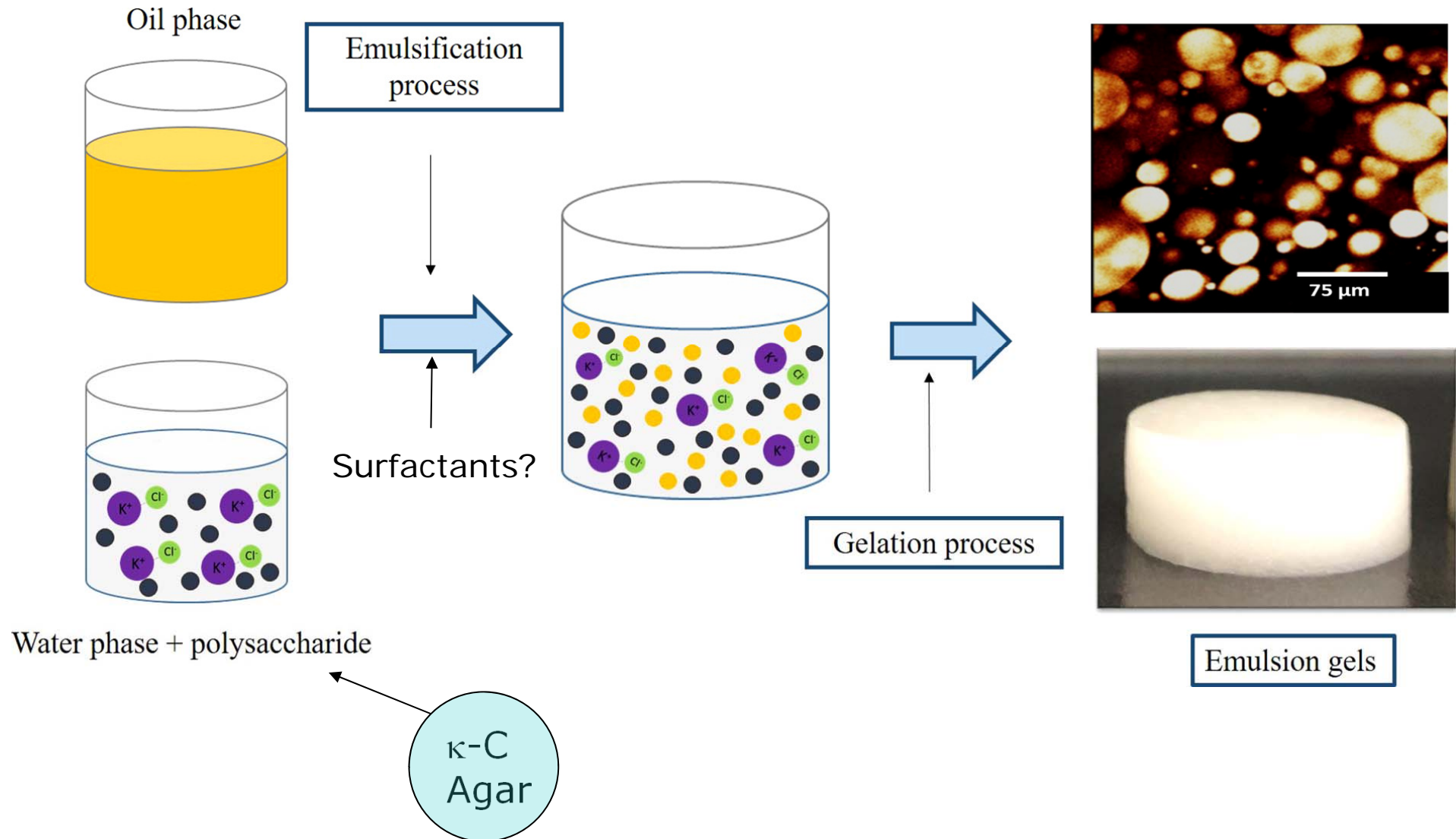


Fontes-Candia et al. *Algal Res.*(2020)

Fontes-Candia et al. *Carb. Polym.*(2021)

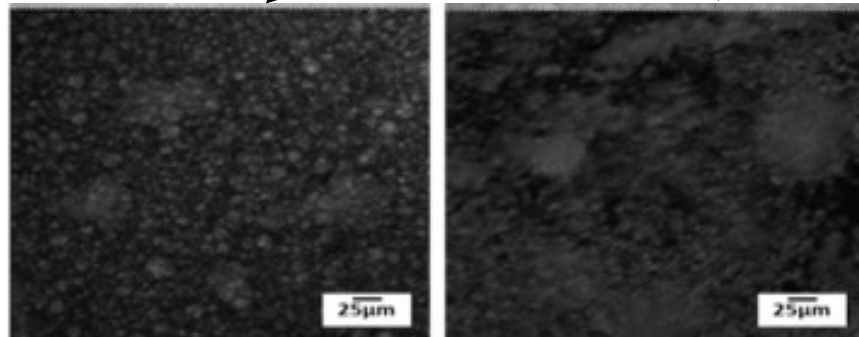
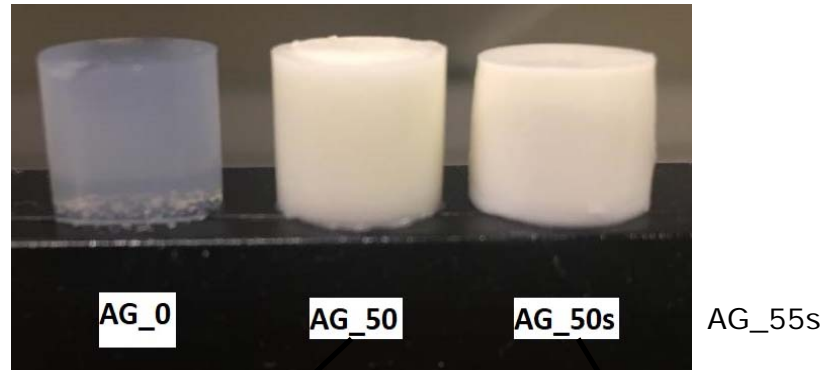
# Emulsion gels

30-50 % w/w

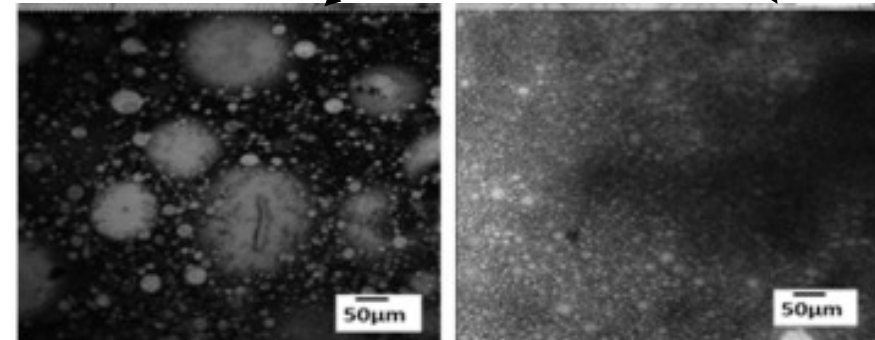
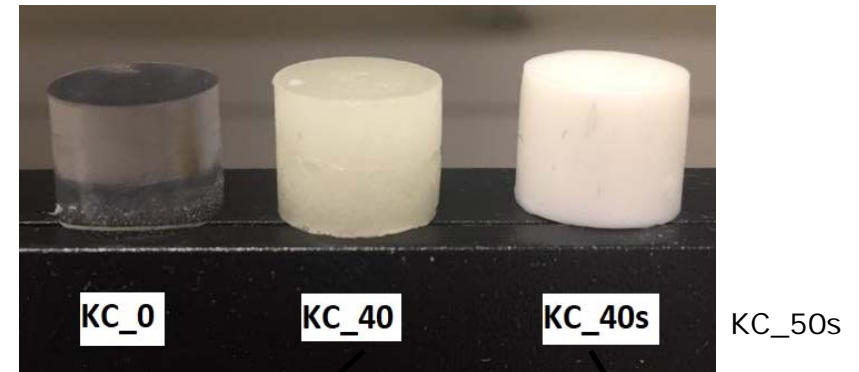


# Emulsion gels

AGAR



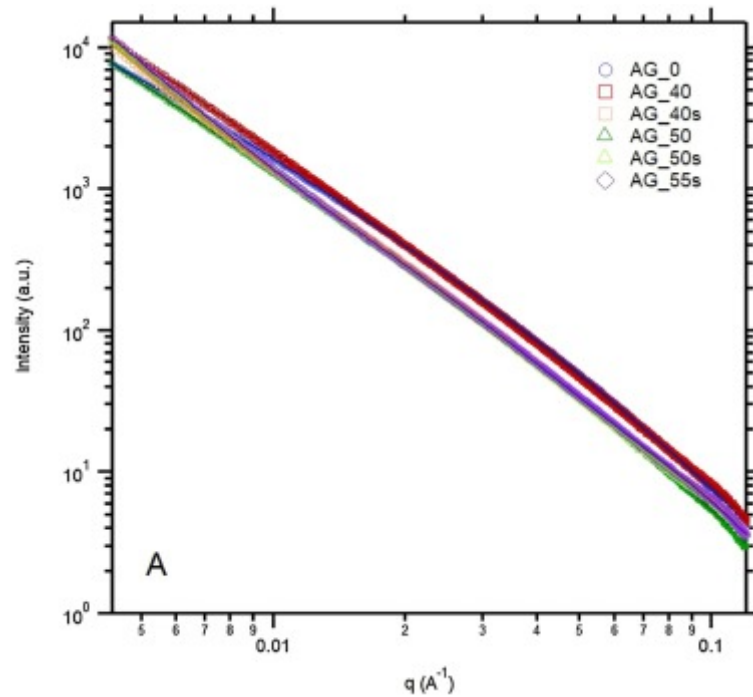
$\kappa$ -C



- Oil droplets distributed within the continuous gelling polysaccharide aqueous phase
- High **emulsifying properties** of **agar** → smaller and more homogeneous droplets
- Competition for the **droplet interface** between surfactants and agar → oil **coalescence**
- Decrease in the viscosity of  $\kappa$ -C with the addition of surfactants → better oil dispersion

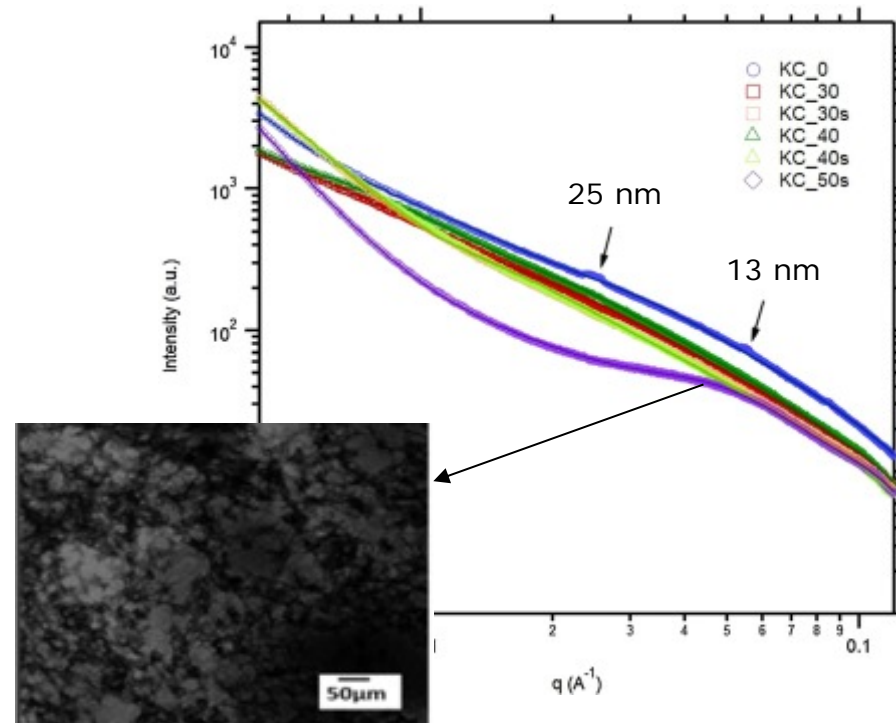
# Emulsion gels

## AGAR



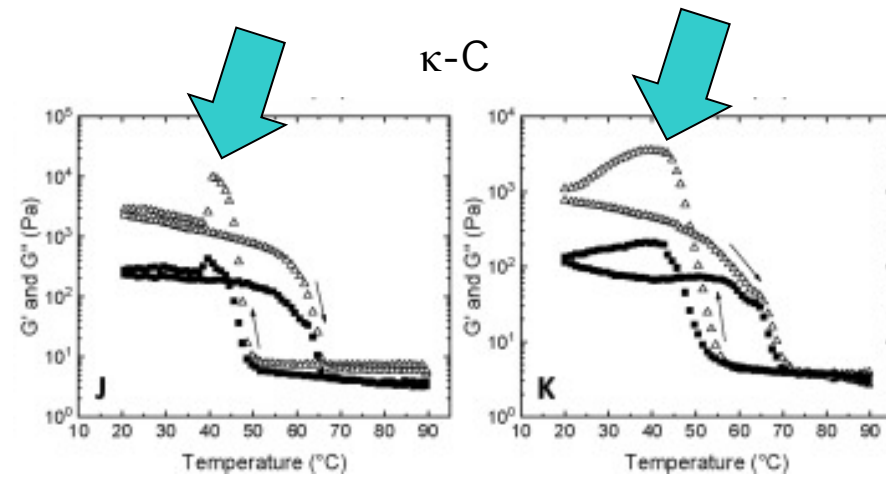
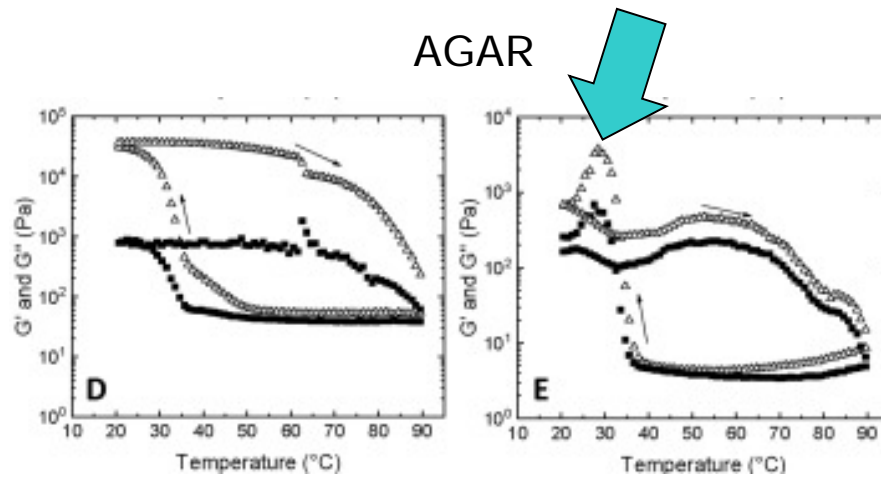
- Branched network structures
- Agar double helices ~ 5–6 nm
- The presence of oil does not have a strong impact on the gel network

## $\kappa$ -C



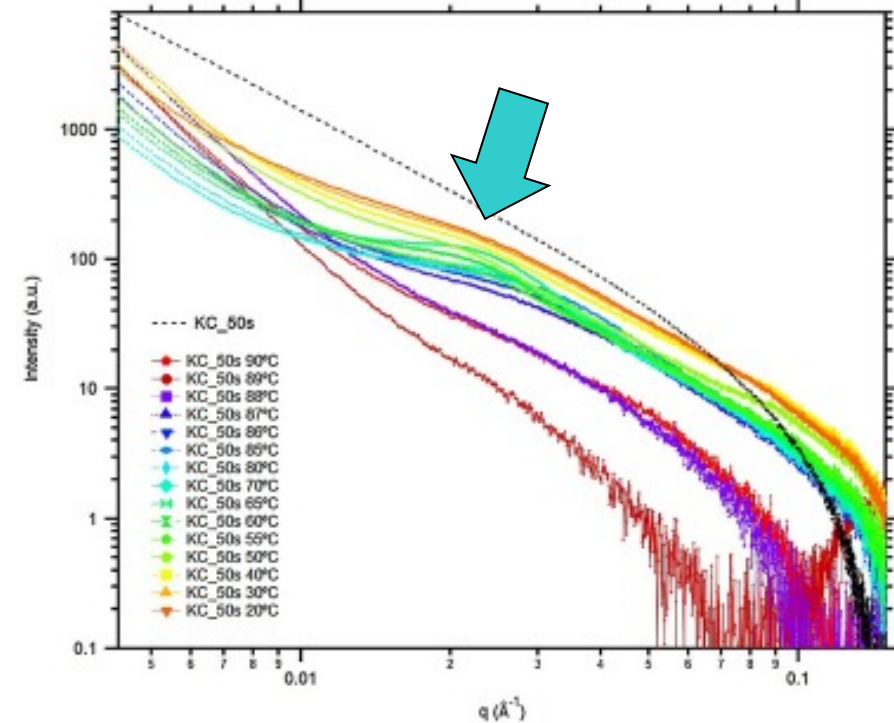
- Less compact network structures
- Less packed  $\kappa$ -C bundles with oil
- $\kappa$ -C double helices ~ 6 nm
- Formation of inverse emulsions?

# Emulsion gels



- Two-phase gelling ( $T_{gel1} \sim 57^\circ\text{C}/T_{gel2} \sim 37^\circ\text{C}$ )  
 → (i) aqueous domain and (ii) agar located at the surface of the oil droplets
- One-phase gelling with surfactant (surfactant located at droplet surface)
- Formation of **metastable intermediate structure** which is disrupted upon further cooling

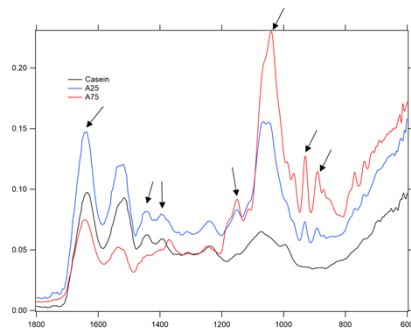
SAXS (cooling ramp)



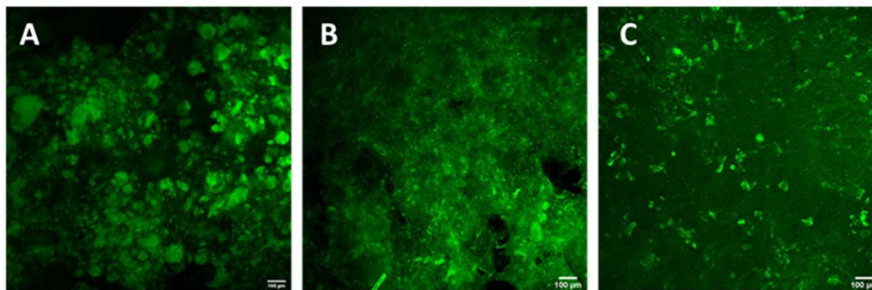
# Ongoing work

## Polysaccharide gels to protect protein products during gastric digestion

Effect of the polysaccharide structure on the protein-polysaccharide interactions



Implications on the digestion process

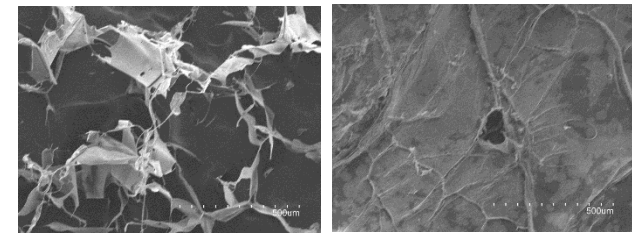


Casein/agar digestion products

Development of different encapsulation structures

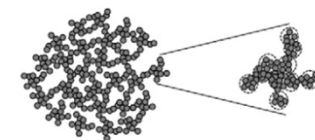
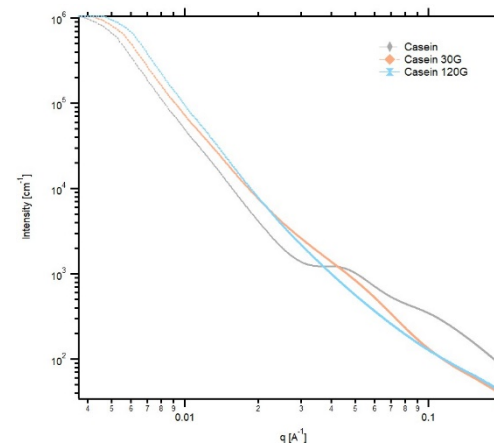


Hydrogels



Casein/polysaccharide aerogels

Structural characterization by SAXS/SANS



Whole cluster, surface fractal structure with  $D^L \sim 3.2$ , low density.

Primary cluster, surface fractal structure with  $D^H \sim 3.2$ , high density.

# Conclusions

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- Scattering techniques are extremely useful to investigate the structure of food polysaccharides in their **native hydrated state**.
- **X-rays and neutrons are complementary** and their combination can provide a more exhaustive structural characterization.
- **Temperature-resolved** scattering **experiments** can be combined with **rheological characterization** to understand the **gelation mechanism** of polysaccharides.
- **In-situ digestion** experiments will be carried out in the future to understand the structural changes undergone by these polysaccharides upon human consumption.

# Acknowledgements



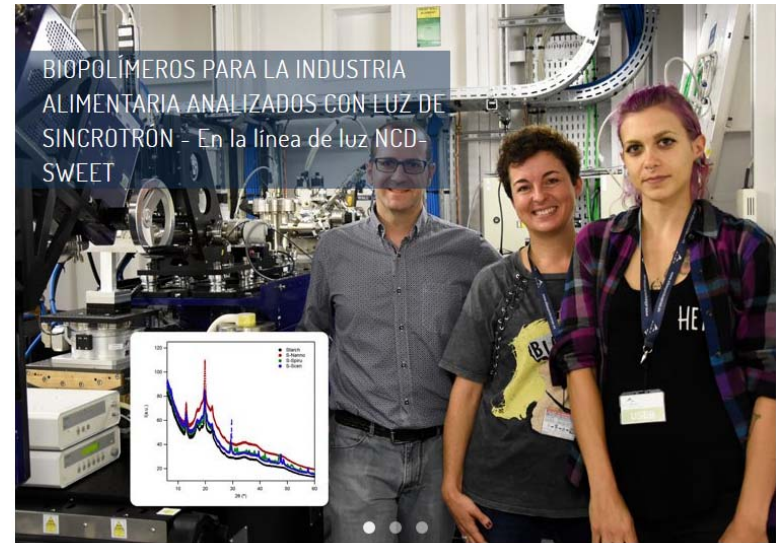
Amparo López-Rubio  
Cynthia Fontes-Candia



Anna Ström  
Patricia López-Sánchez



Juan Carlos Martínez



Elliot Gilbert  
Christine Rehm  
Liliana de Campo  
Anna Sokolova

