



Towards Correlative X-Ray Tomographic Imaging of Membranes for Improving Food Processing

G. Rudolph, J. Thuvander, E. Larsson, S. Hall, T. Nilsson Pingel,
Pablo Villanueva-Perez, F. Lipnizki



Outline

- Membrane processes
 - Advantages and challenges in food processing
 - Definition of membrane fouling
- Towards correlative X-Ray tomographic imaging
- Take home messages



Advantages of membrane processes for food processing



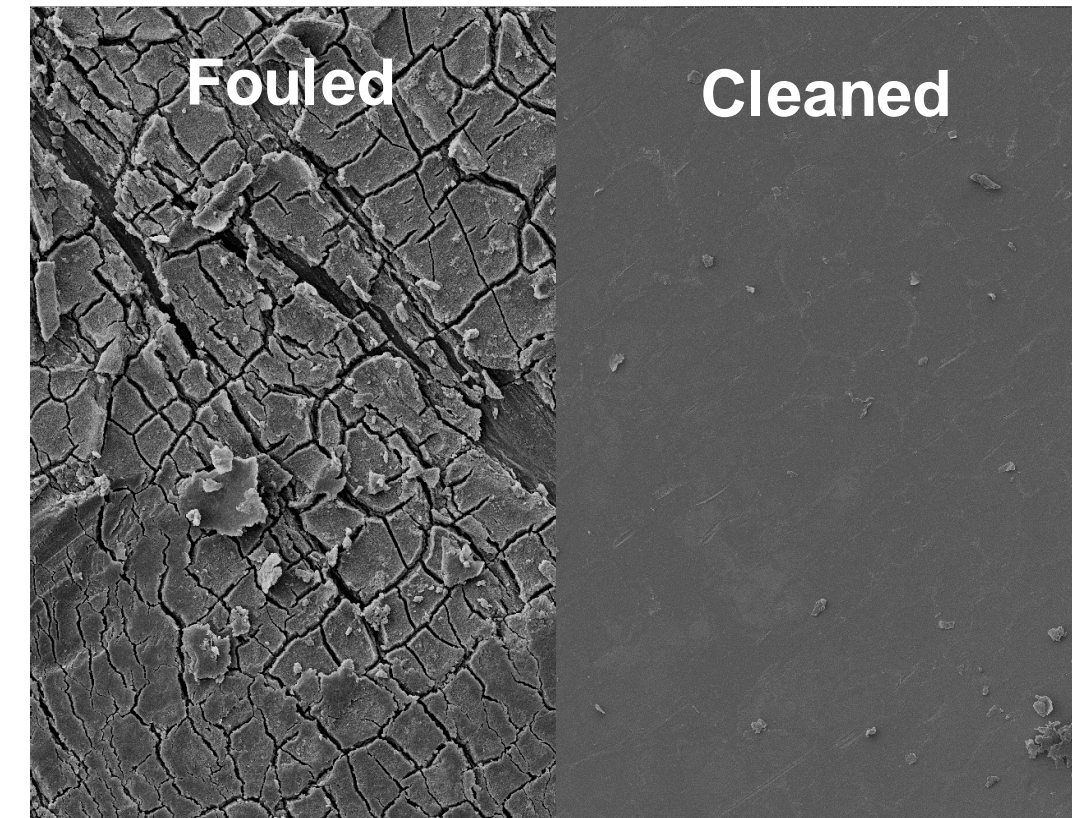
- Gentle product treatment due to moderate temperature changes during processing.
- High selectivity based on unique separation mechanism
e.g. sieving, solution-diffusion or ion-exchange mechanism.
- Compact and modular design for ease of installation and extension.
- Low energy consumption compared to condensers and evaporators.
- Integration with other separation technologies in synergy or hybrid processes.



Challenges of membrane processes for food processing

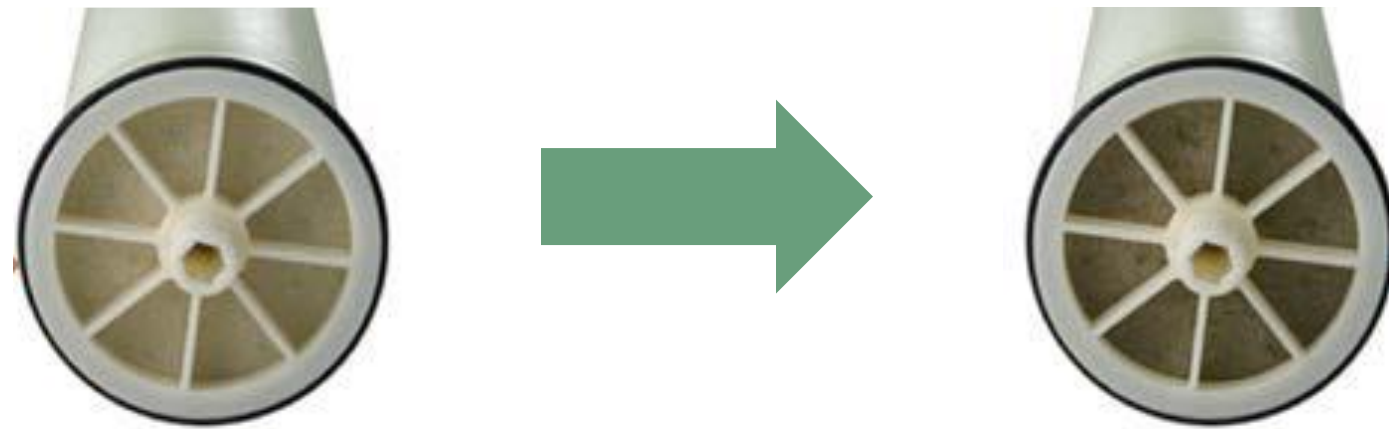


- Short membrane life cycle and high membrane costs.
- Products have to be liquids or gaseous/vapour.
- Membrane performance is often limited by temperature, pH and chemical resistance.
- Limited economy of scale – membrane plants scale linearly.
- Tendency to **membrane fouling** and thus need for **membrane cleaning**.



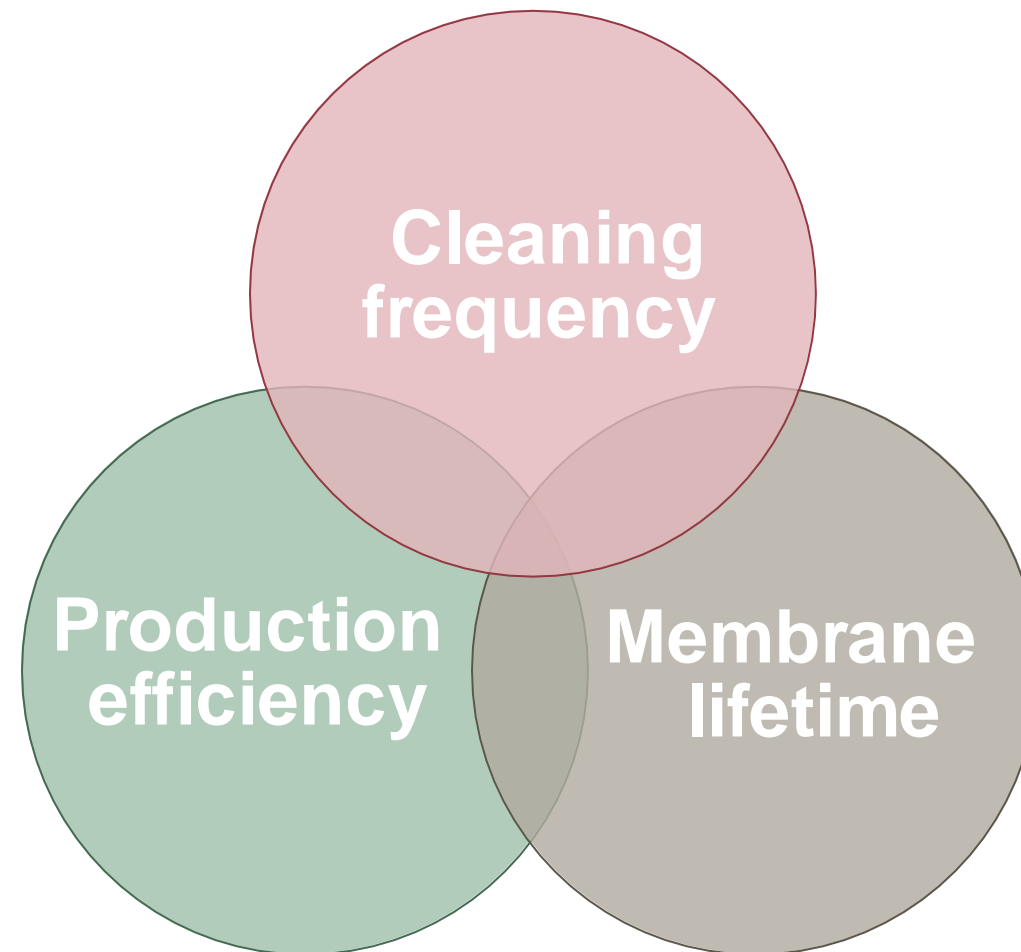
Membrane fouling?

“ Process resulting in loss of performance of a membrane due to the deposition of suspended or dissolved substances on its external surfaces, at its pore opening, or within its pores ”



Fouling and cleaning is costly

Production time losses due to cleaning and membrane replacement



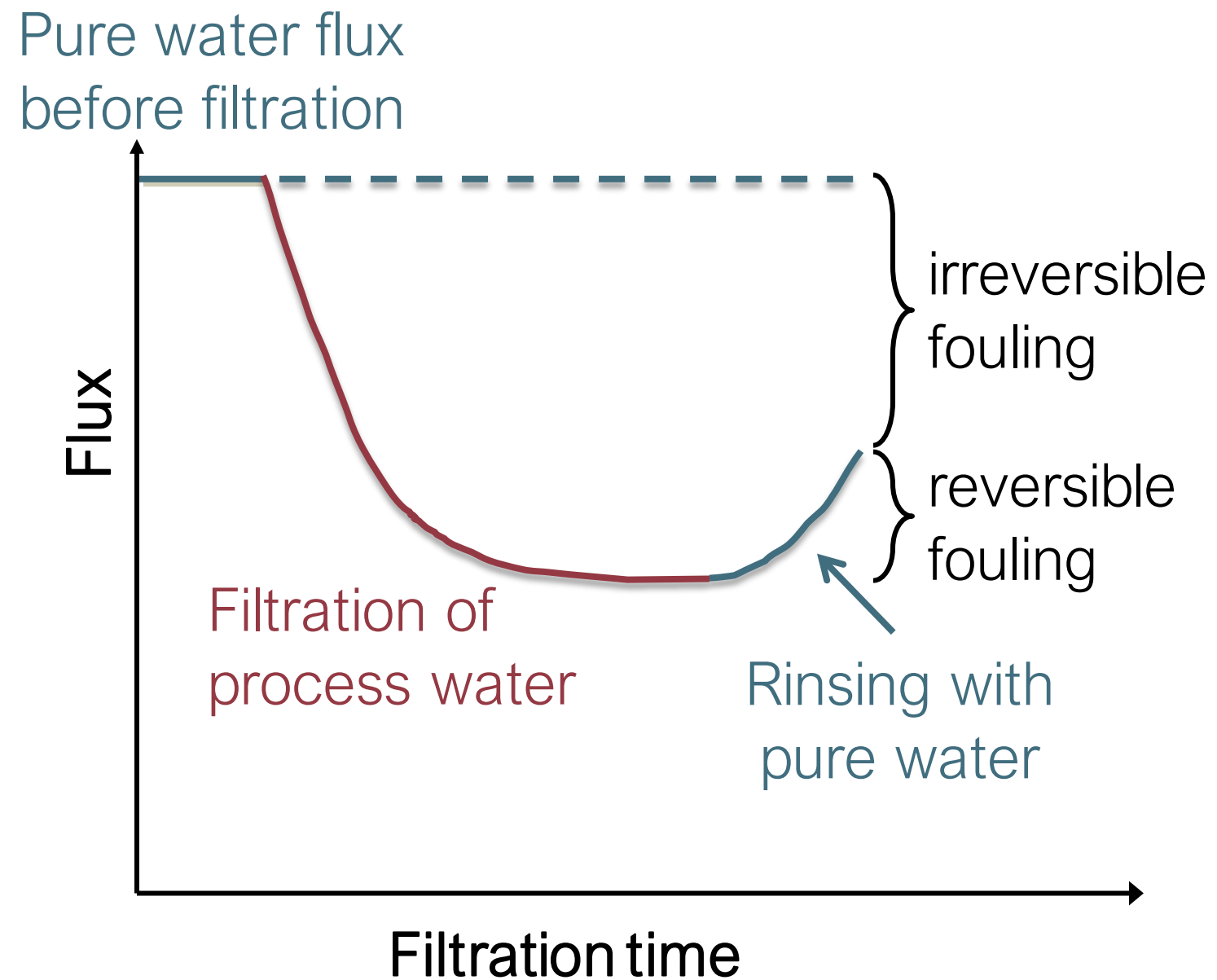
Irreversible fouling and cleaning cycles reduce membrane lifetime

Total daily plant capacity has to be increased as a consequence of cleaning

Annual costs

- Membrane cleaning: 5-20 % of CAPEX
- Membrane replacement: 2-5 % of CAPEX

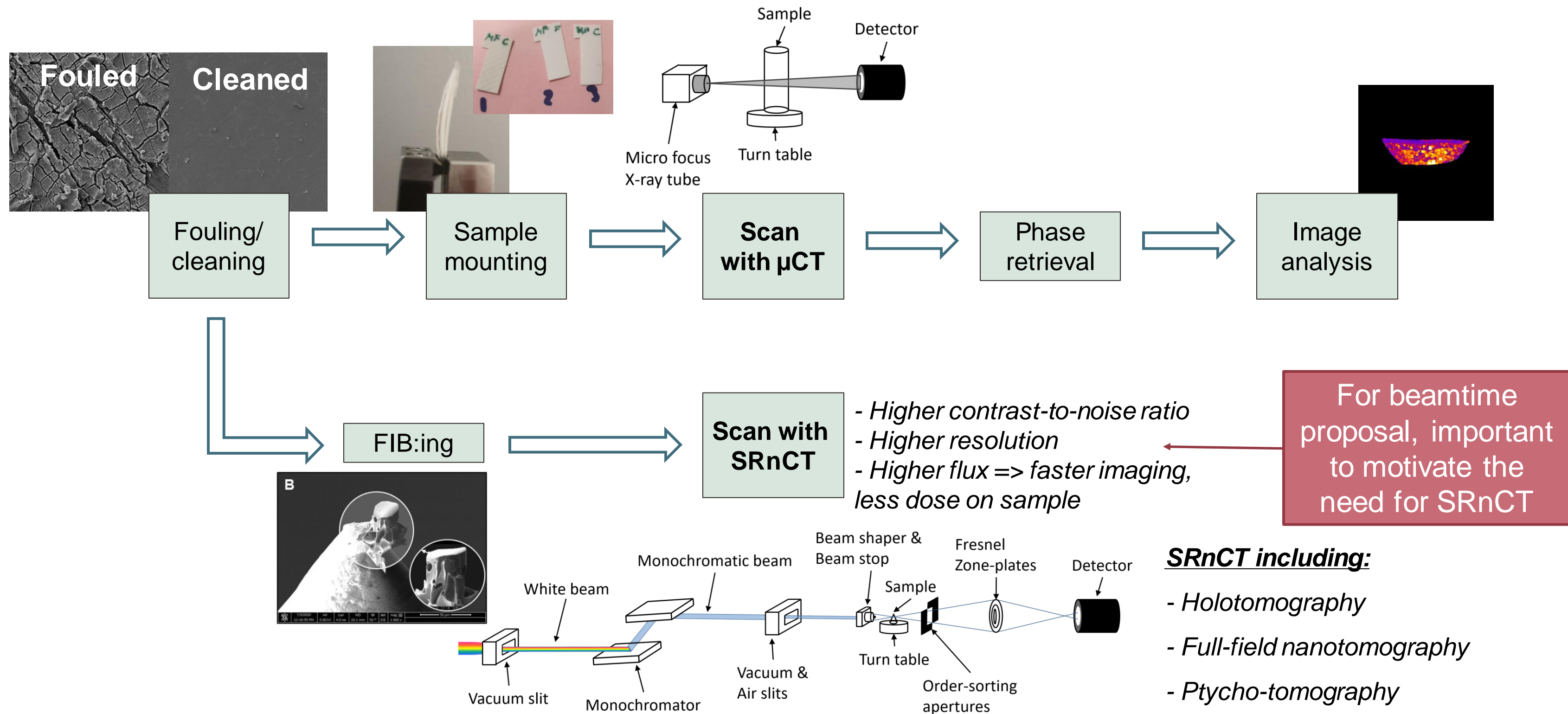
Membrane fouling?



- No information about the fouling layer structure
- No information about membrane altering due to fouling and cleaning



How we have worked with correlative X-ray tomography



Toolkits for teaching tomography

Additional lectures on:

- Small Angle X-ray Scattering (SAXS), Shun Yu, RISE
- High resolution coherent X-ray imaging, Pablo Villanueva-Perez, LU

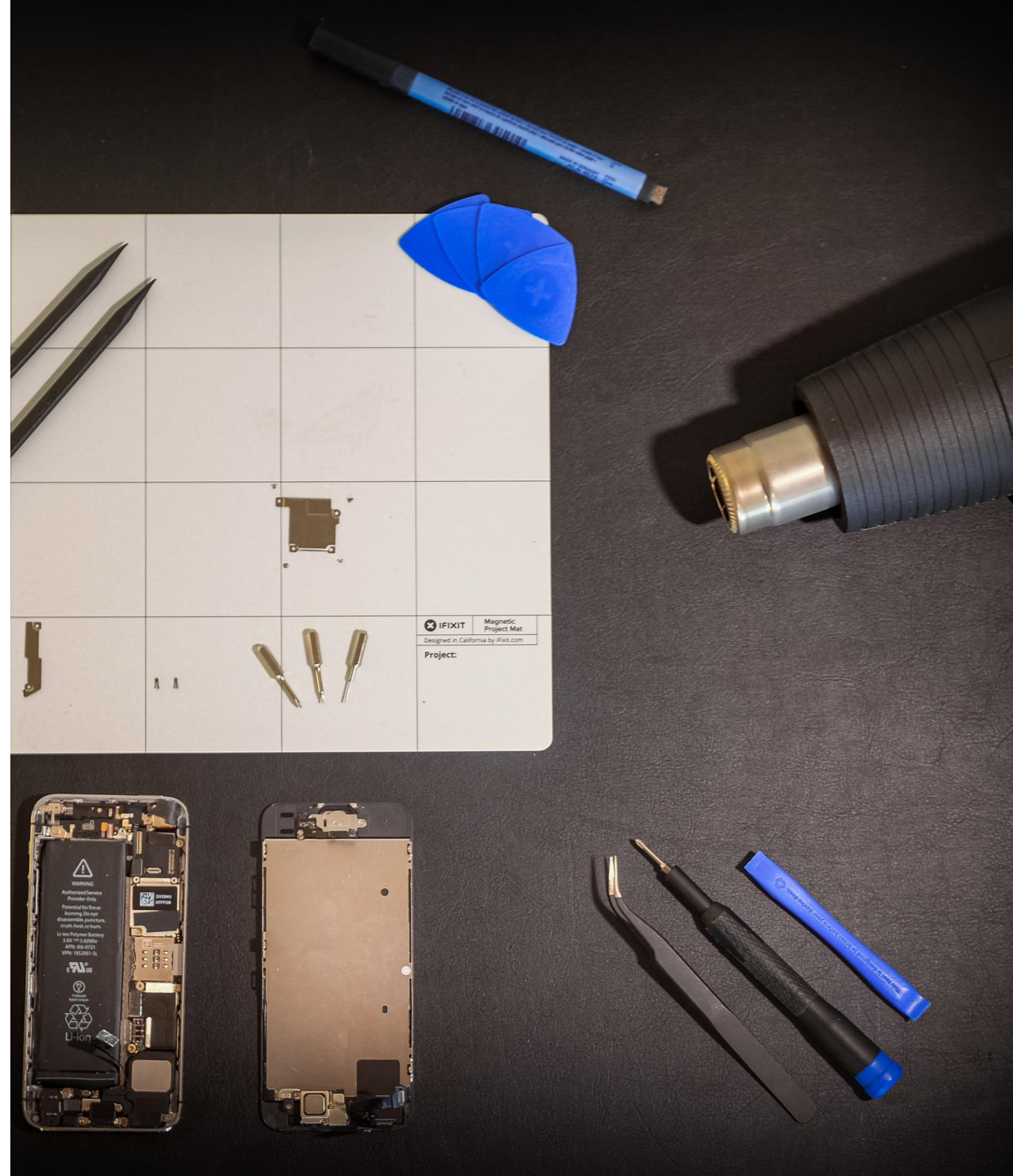


LUND
UNIVERSITY



Call: Increasing capacity and skills of PhD students regarding industrially relevant neutron and synchrotron-based analytical methods – 2019

Project: Measurements of membrane fouling in lignocellulosic biorefineries by ptycho-tomography, 2019-03613



Kitchen-Based Light Tomography – a home-based educational scanner for training the next generation of X-ray and Neutron tomography users

Emanuel Larsson¹ & Stephen A. Hall¹

¹Division of Solid Mechanics, Lund University, Lund, Sweden



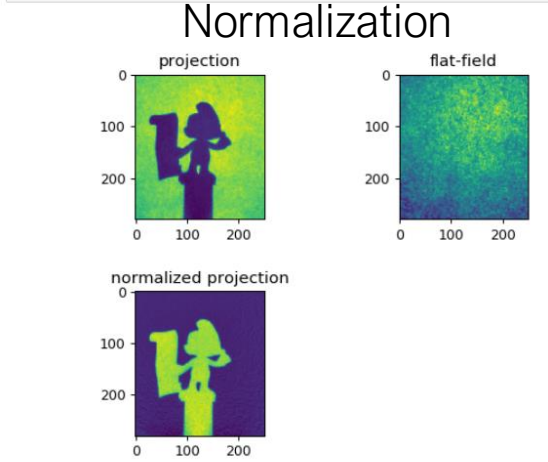
<https://www.blue-scientific.com/tosca-2020/>

Reconstruction – using TomoPy

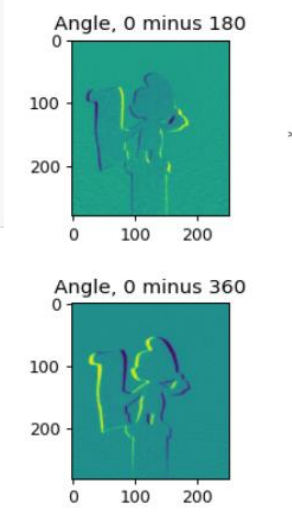
```

matplotlib notebook

plt.figure()
prj = plt.figure()
prj.subplots_adjust(hspace=5)
ax = prj.add_subplot(2, 2, 1)
ax.set_title('projection')
ax.imshow(tomo[0])
ax2 = prj.add_subplot(2, 2, 2, sharex=ax, sharey=ax)
ax2.set_title('flat-field')
ax2.imshow(flat[0])
ax3 = prj.add_subplot(2, 2, 3, sharex=ax, sharey=ax)
ax3.set_title('normalized projection')
ax3.imshow(data[0])
plt.show()
    
```



Tilt correction



3D Reconstruction

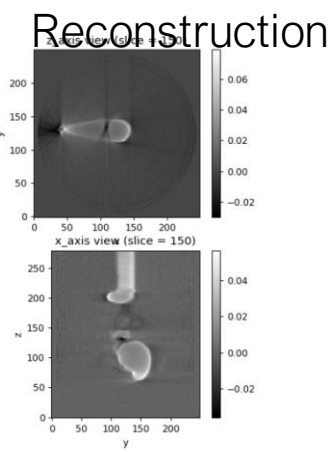
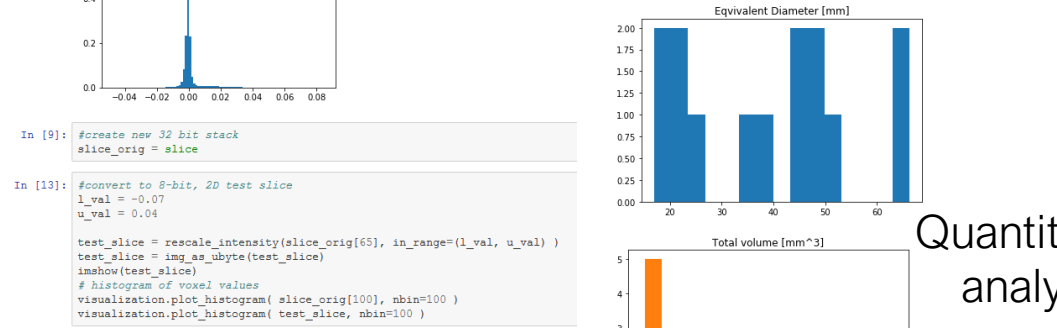


Image Processing and Quantitative Analysis

```

In [8]: # histogram of voxel values
visualization.plot_histogram( slice, nbins=100 )

diameter = np.array([])
tot_vol = np.array([])
for i in range(no_inclus):
    diameter = np.append(diameter, props[i].equivalent_diameter)
    tot_vol = np.append(tot_vol, props[i].area * voxel_volume)
parameter_list = [diameter, tot_vol]
title_list = ['Equivalent Diameter [mm]', 'Total volume [mm^3]']
for i in range(len(parameter_list)):
    plt.hist(parameter_list[i], bins=15)
    plt.title(title_list[i])
    plt.show(i)
    
```



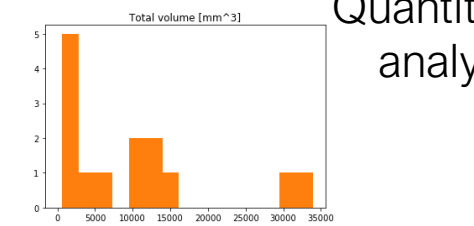
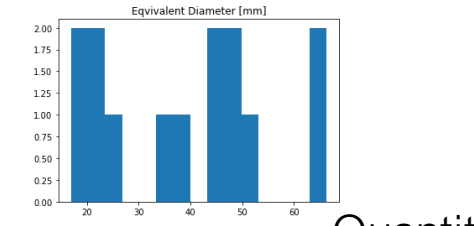
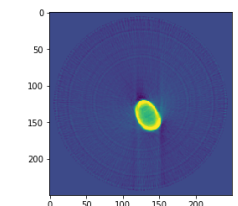
32 to 8 bit conversion

```

In [9]: #create new 32 bit stack
slice_orig = slice

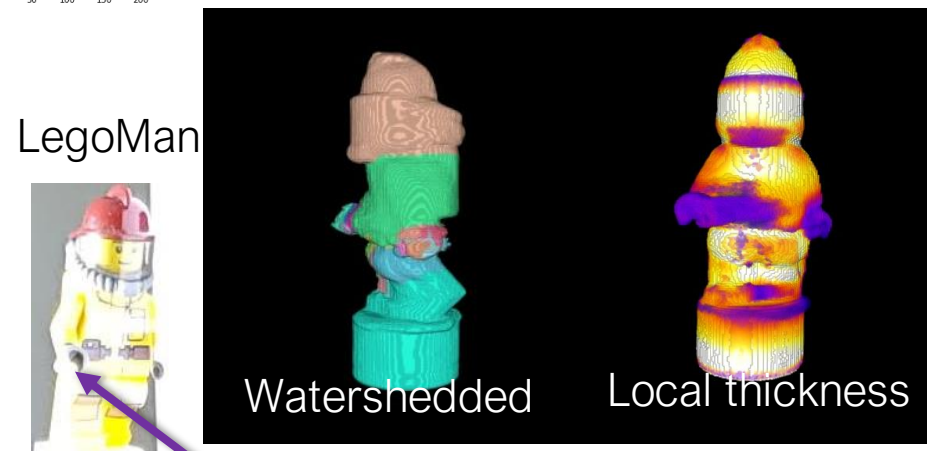
In [13]: #convert to 8-bit, 2D test slice
l_val = -0.07
u_val = 0.04

test_slice = rescale_intensity(slice_orig[65], in_range=(l_val, u_val))
test_slice = img_as_byte(test_slice)
imshow(test_slice)
# histogram of voxel values
visualization.plot_histogram( slice_orig[100], nbins=100 )
visualization.plot_histogram( test_slice, nbins=100 )
    
```



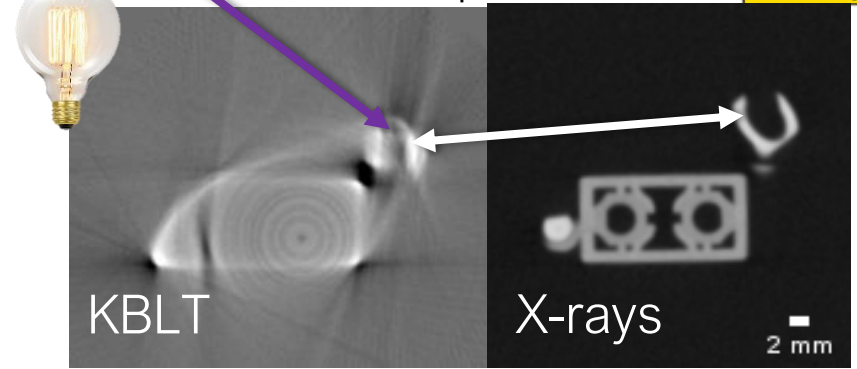
Quantitative analysis

3D-rendering



Comparison

X-ray

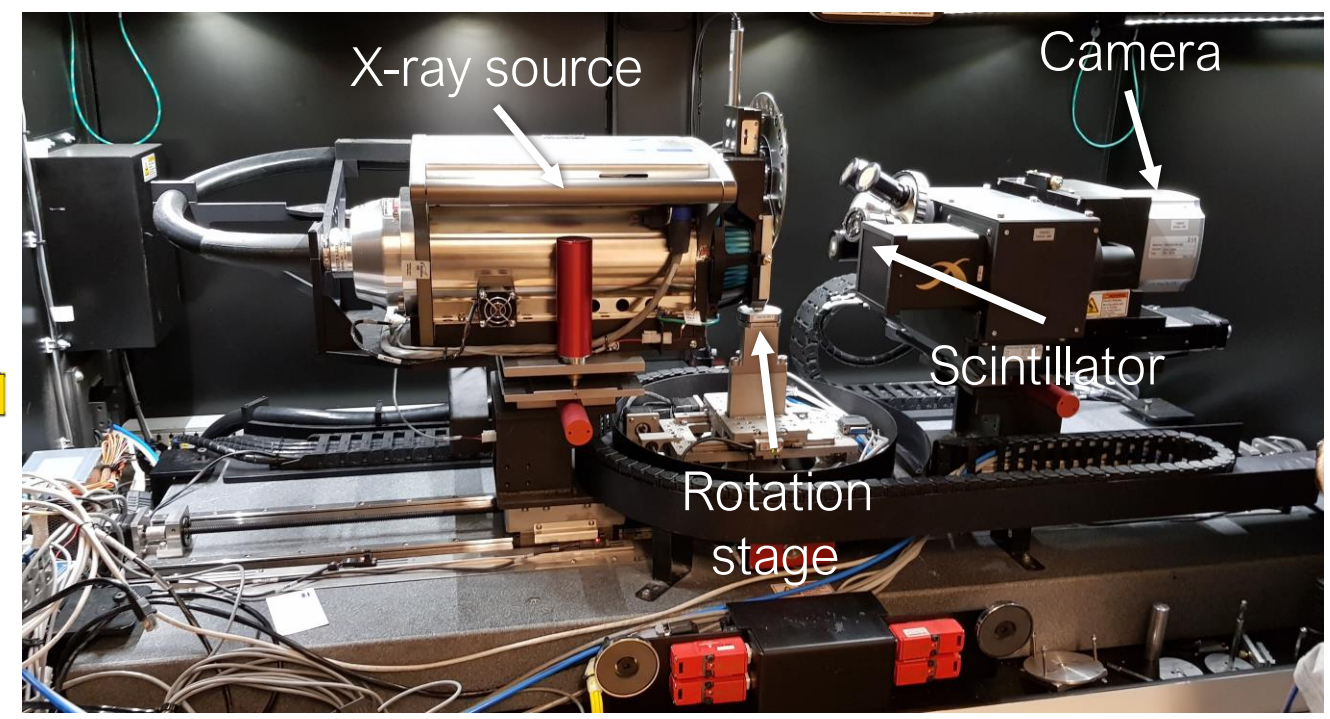
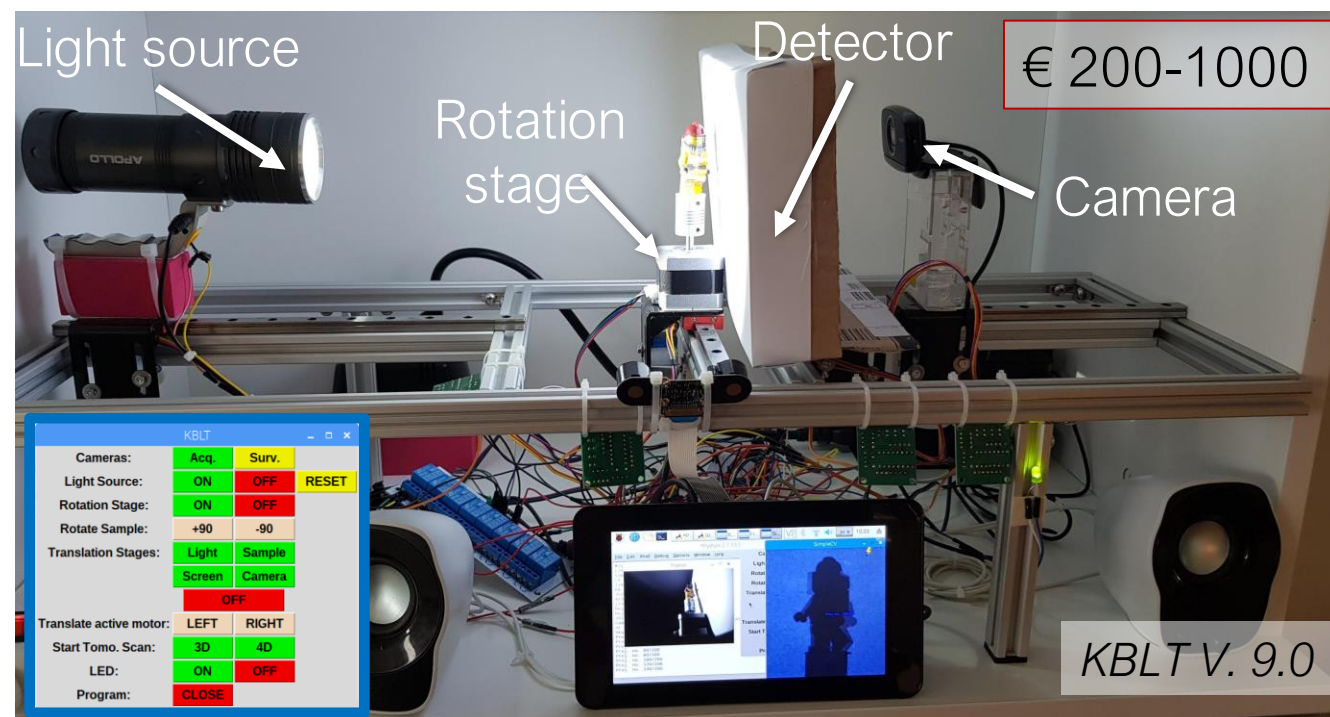


KBLT

X-rays

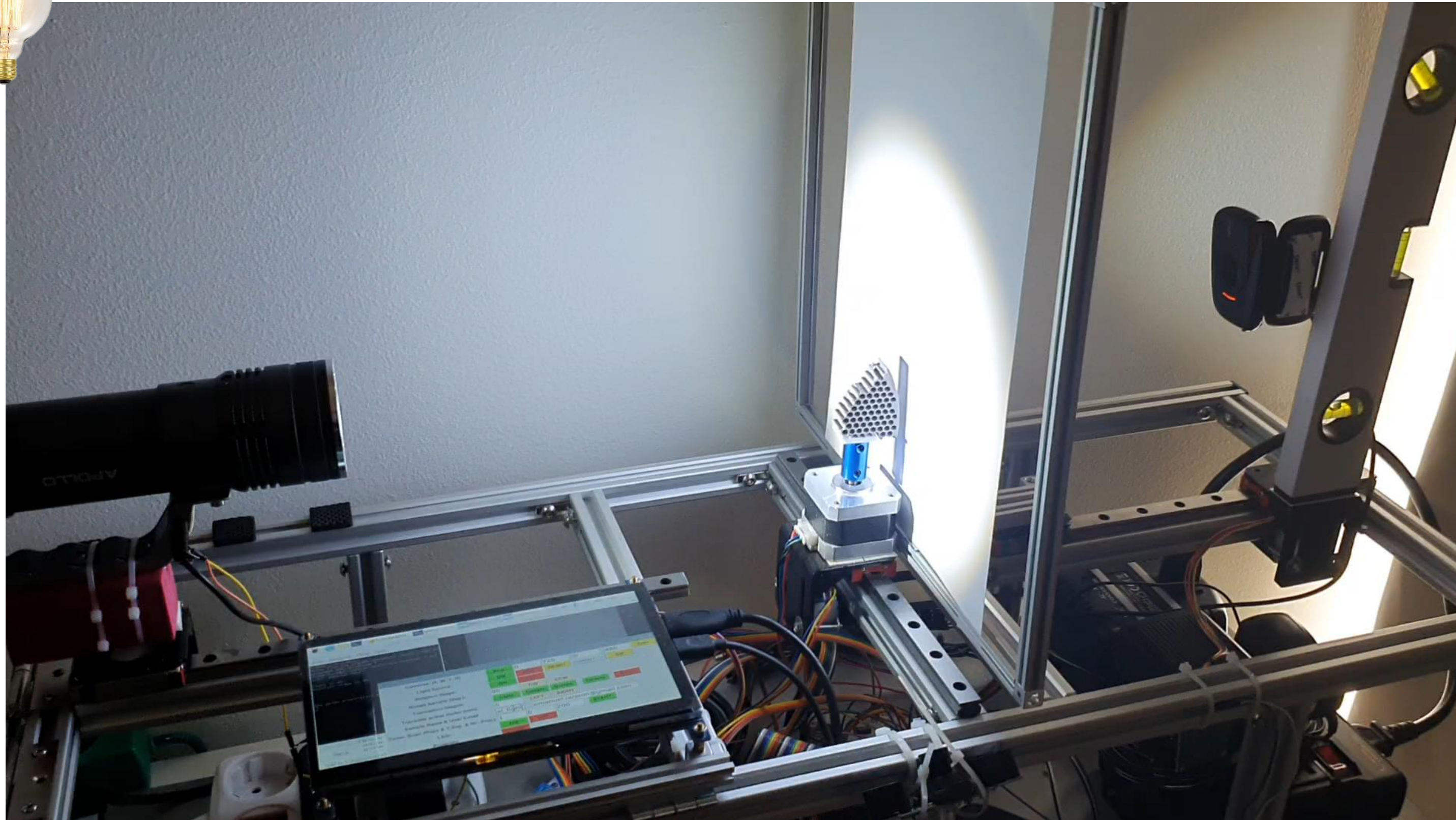
2 mm

Has been used to train both PhD students and Post Docs.

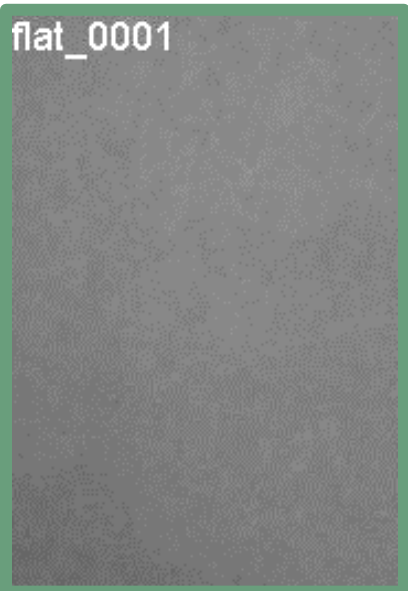
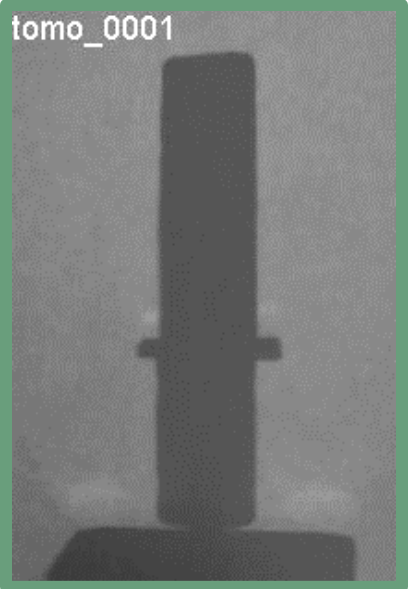


Zeiss Xradia 520 Versa at 4D Imaging Lab at Lund University.

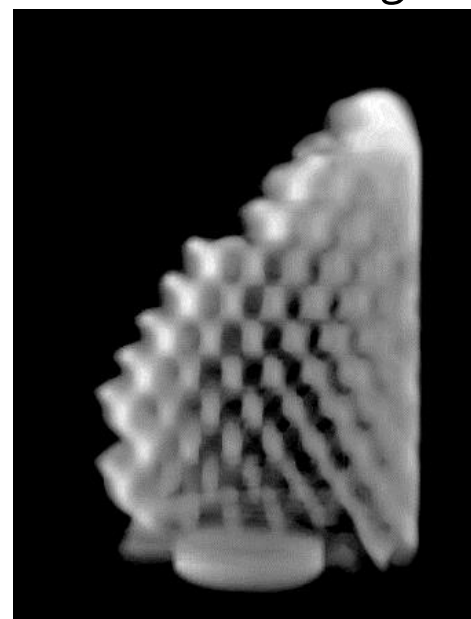
KBLT v.14.0 - pre-recorded demo!



Camera view

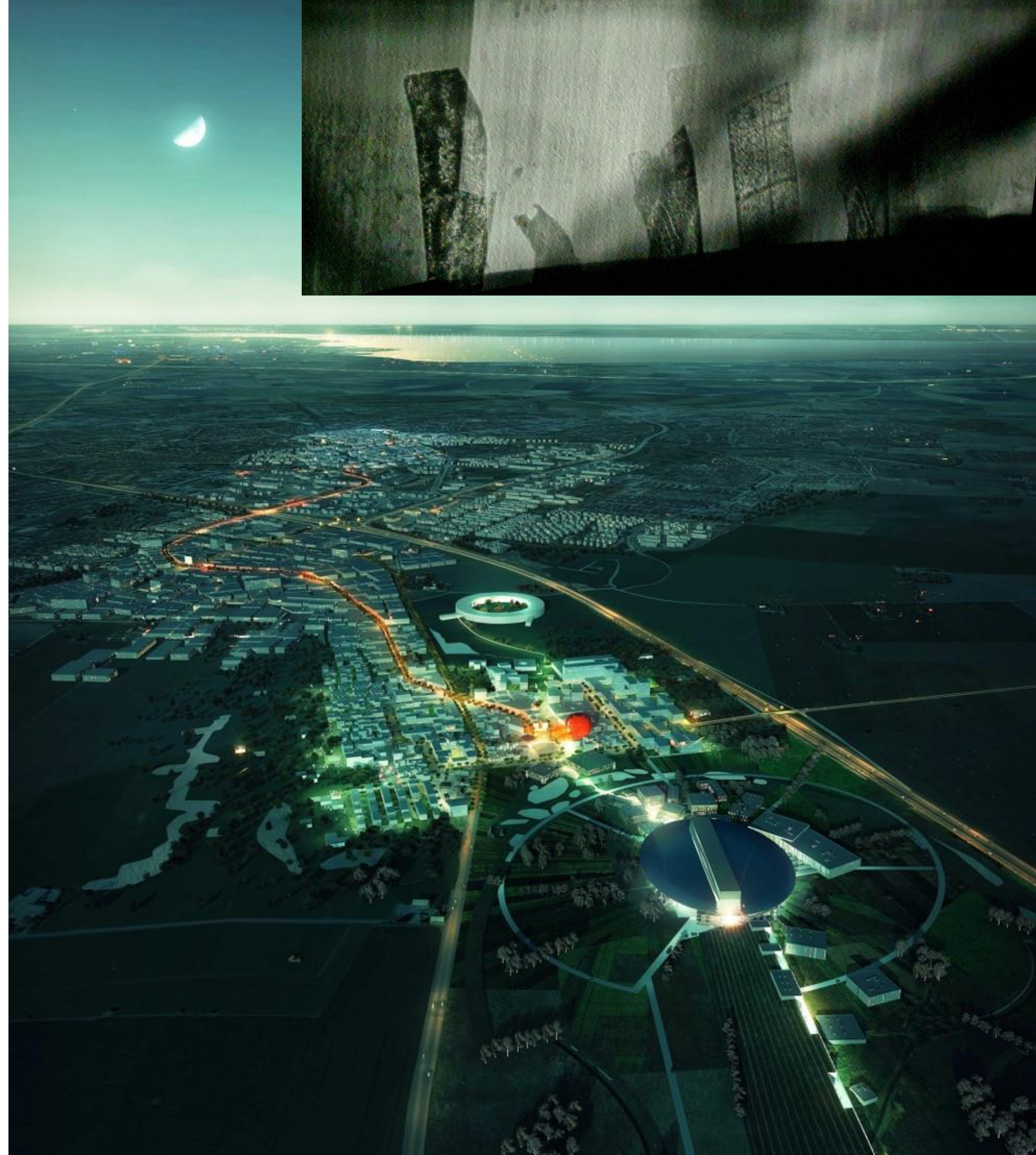


3D-rendering



Membrane sample provided by Rudolph Gregor, Lund University, Sweden.

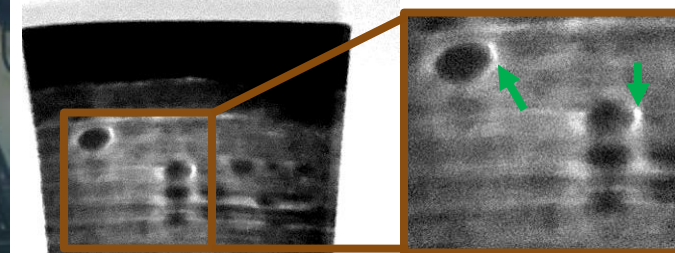
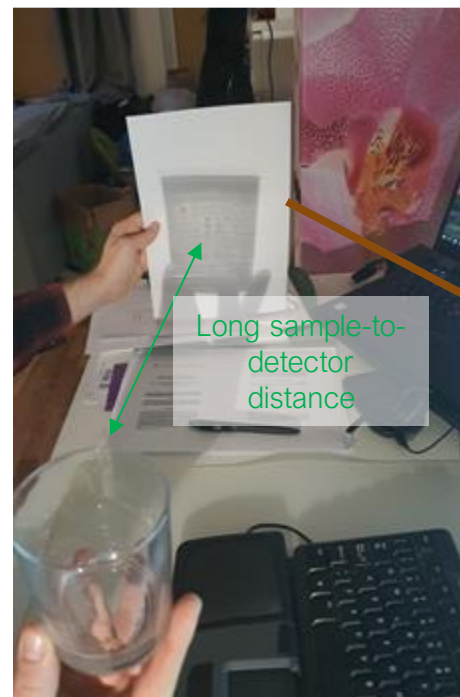
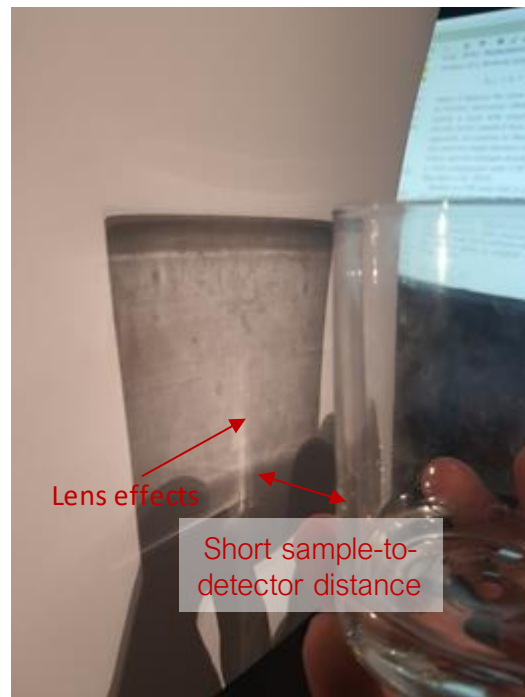
Imaging techniques
with normal light vs.
X-rays





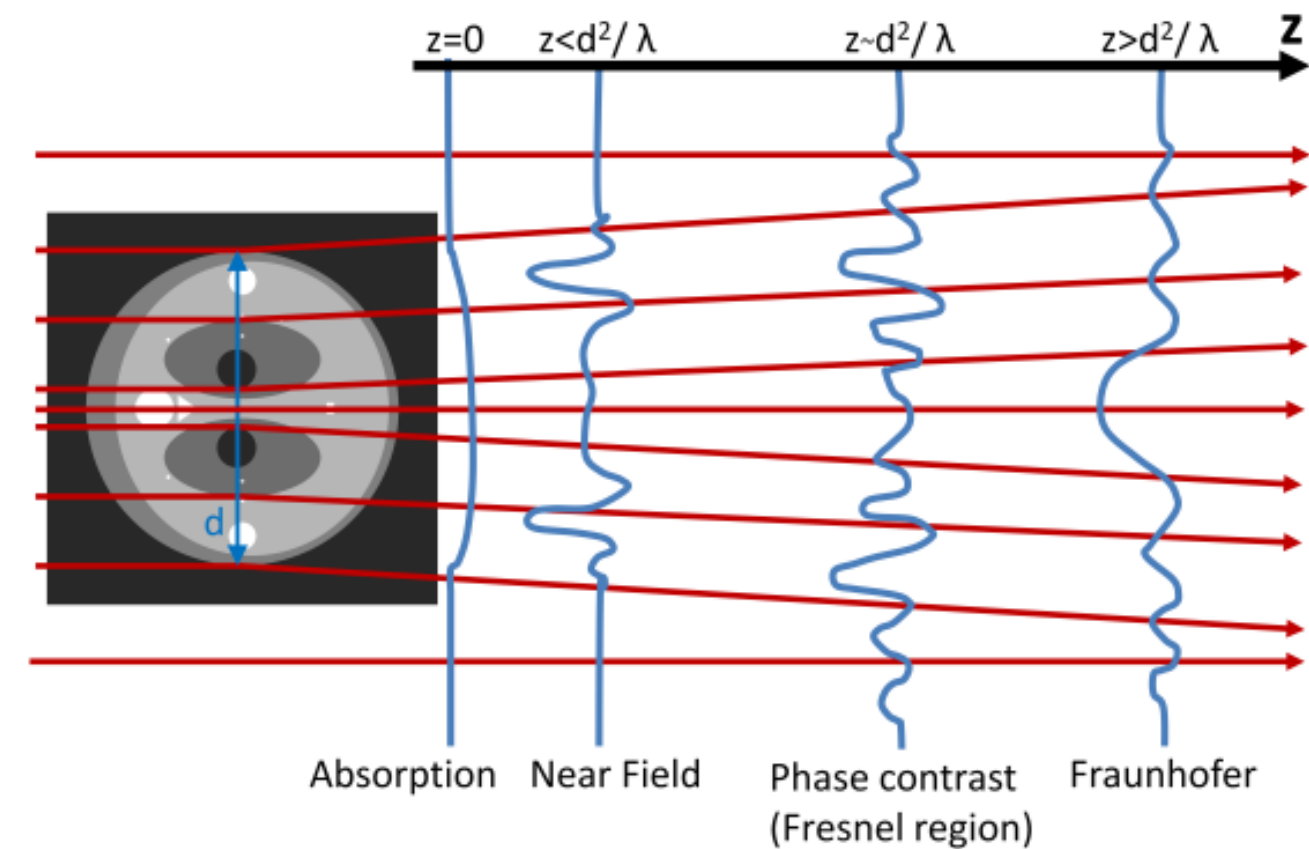
To reconstruction 'diffuse' patterns there is the need to perform so called 'phase retrieval', in order to decouple phase shift from absorption.

**INCREASING RESOLUTION/
SAMPLE-TO-DETECTOR DISTANCE**



Absorption contrast

Phase contrast

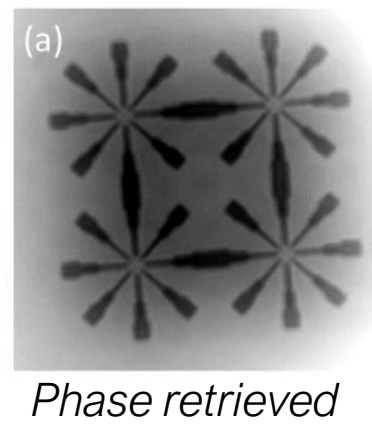


X-ray

Full-field Synchrotron X-ray holographic nanotomography

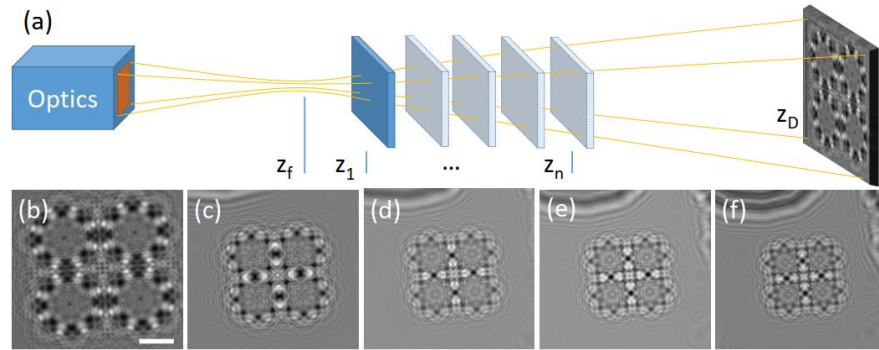
- Absorption
- Phase Contrast

- 1) Lensless
- 2) Zone plate



2D test pattern

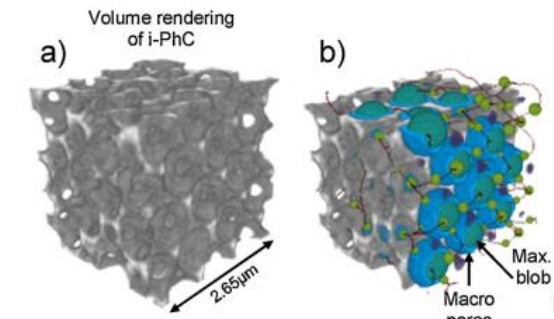
Typical Resolution: $>100\text{ nm}$



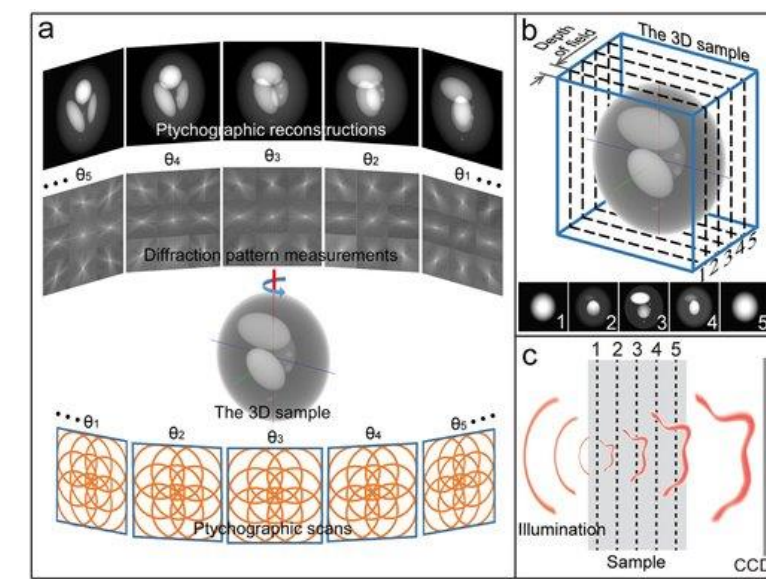
Langer, M., Zhang, Y., Figueirinhas, D., Forien, J. B., Mouton, C., Mokso, R., & Villanueva-Perez, P. (2020). PyPhase--a Python package for X-ray phase imaging. *arXiv preprint arXiv:2012.07942*.

Kalbfleisch S., Zhang Y., Kahnt M., Buakor K., Langer M., Dreier T., Dierks H., Stjarneblad P., **Larsson E.**, Gordeyeva K., Chayanun L., Soederberg D., Wallentin J., Bech M. and Villanueva-Perez P., X-ray in-line holography and holotomography at the NanoMAX beamline, *Submitted to the Journal of Synchrotron Radiation* 2021.04.19

Typical Resolution: $>16\text{ nm}$



Furlan, K. P., **Larsson, E.**, Diaz, A., Holler, M., Krekeler, T., Ritter, M., ... & Janßen, R. (2018). Photonic materials for high-temperature applications: Synthesis and characterization by X-ray ptychographic tomography. *Applied materials today*, 13, 359-369.



Li, P., & Maiden, A. (2018). Multi-slice ptychographic tomography. *Scientific reports*, 8(1), 1-10.

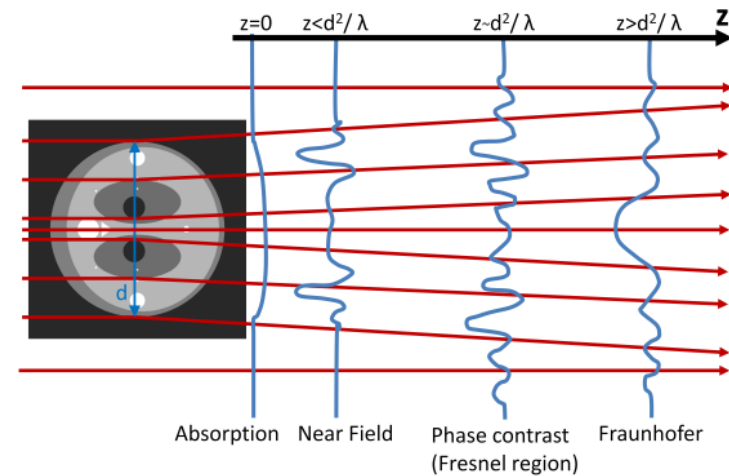
Synchrotron X-ray ptycho-nanotomography

- Absorption
- Phase Contrast

INCREASING RESOLUTION/
SAMPLE-TO-DETECTOR DISTANCE

Full-field lab-based & Synchrotron X-ray Microtomography

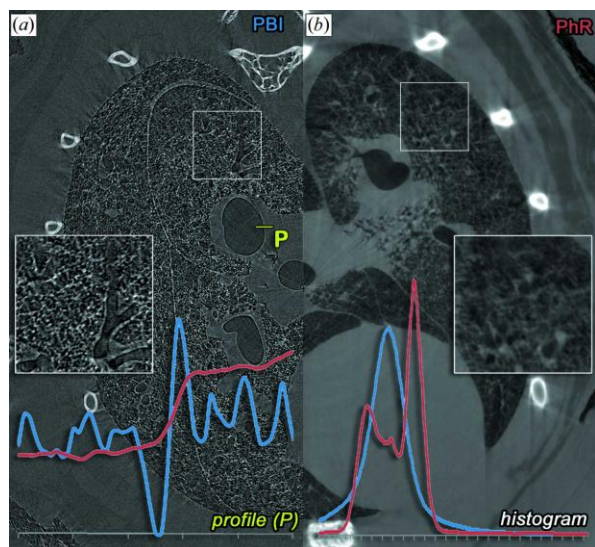
- Absorption
- In-line Phase Contrast



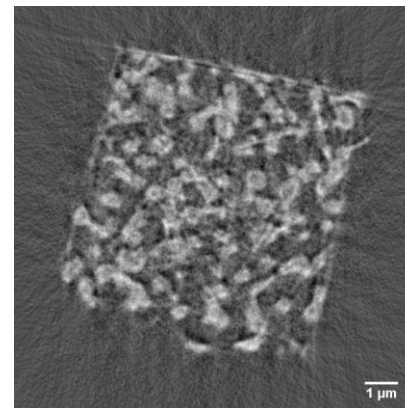
Mouse lung

Typical Resolution: $>0.7\ \mu\text{m}$

Mohammadi, S., **Larsson, E.**, Alves, F., Dal Monego, S., Biffi, S., Garrovo, C., ... & Dullin, C. (2014). Quantitative evaluation of a single-distance phase-retrieval method applied on in-line phase-contrast images of a mouse lung. *Journal of synchrotron radiation*, 21(4), 784-789.



Larsson, E., Gürsoy, D., De Carlo, F., Lilleodden, E., Storm, M., Wilde, F., ... & Greving, I. (2019). Nanoporous gold: a hierarchical and multiscale 3D test pattern for characterizing X-ray nanotomography systems. *Journal of synchrotron radiation*, 26(1), 194-204.



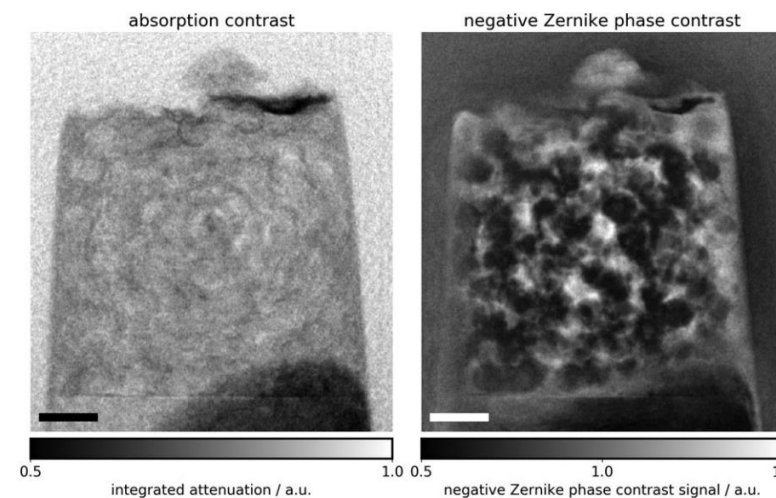
Nanoporous gold

Storm, M., Döring, F., Marathe, S., David, C., & Rau, C. (2020). The Diamond I13 full-field transmission X-ray microscope: a Zernike phase-contrast setup for material sciences. *Powder Diffraction*, 1-7.

Full-field Synchrotron X-ray nanotomography

- Absorption
- Zernike Phase Contrast

Typical Resolution: $>60\text{ nm}$



Photonic glass

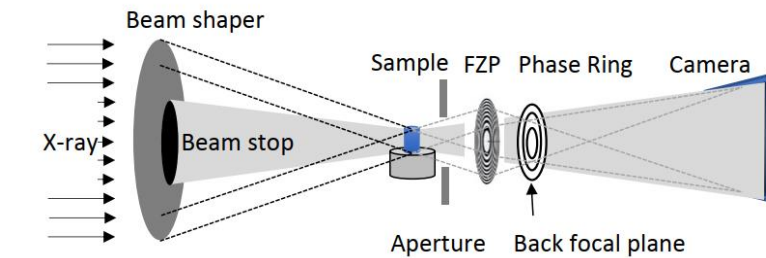
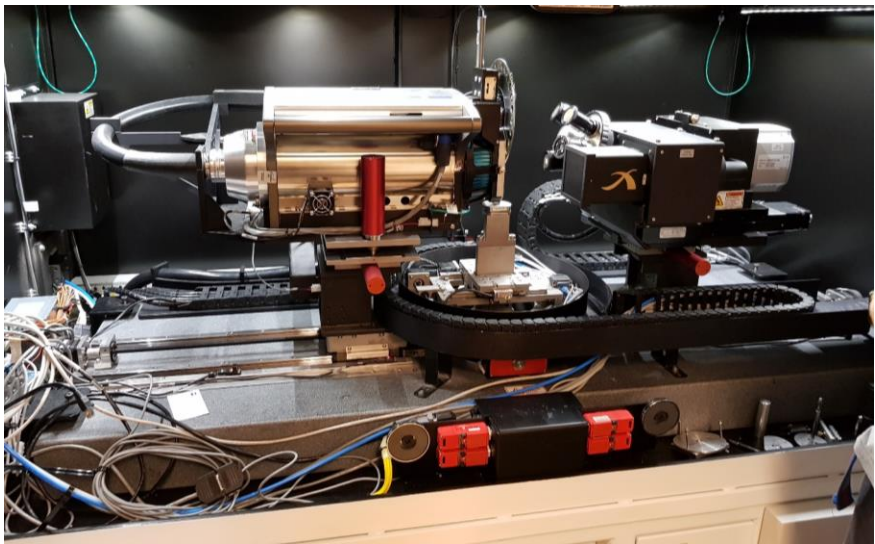


Figure 1. Schematic of the ZPC setup at the imaging beamline P05.

Flenner, S., **Larsson, E.**, Furlan, K., Laipple, D., Storm, M., Wilde, F., ... & Greving, I. (2018). Nanotomography of inverse photonic crystals using zernike phase contrast. *Microscopy and microanalysis*, 24(S2), 146-147.



From microtomography towards ptycho-tomography of membrane fouling in lignocellulosic biorefineries

Beamtime proposals sent in to:

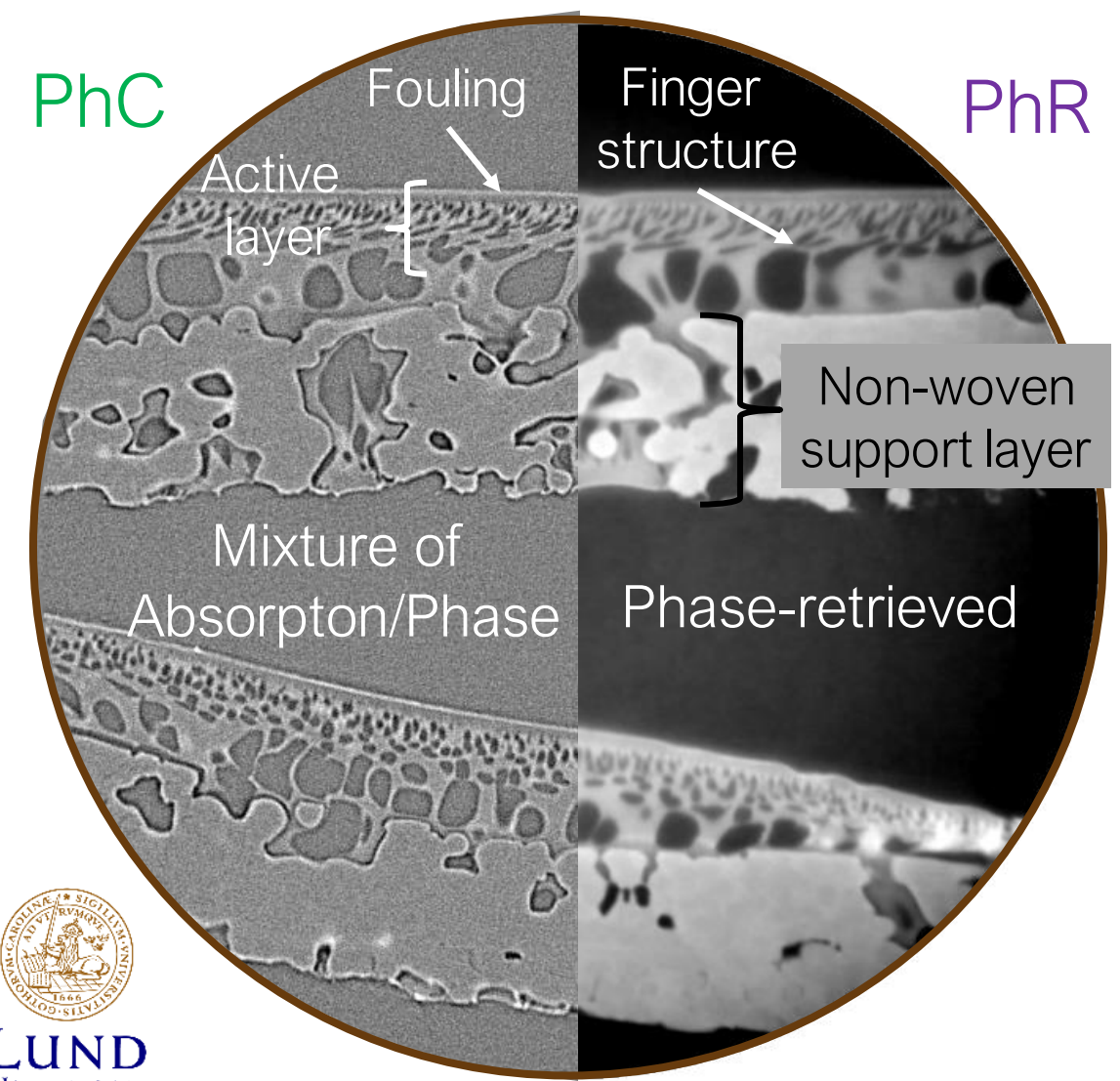
- NanoMAX, in total 2 during 2020-2021 - declined
- cSAXS, PSI, in total 2 during 2020-2021 (on waiting list)

4D Imaging Lab, Division of Solid Mechanics, Faculty of Engineering, Lund University
<https://www.solid.lth.se/resources/4d-imaging-lab/>



Center for Quantification of Imaging Data from MAX IV

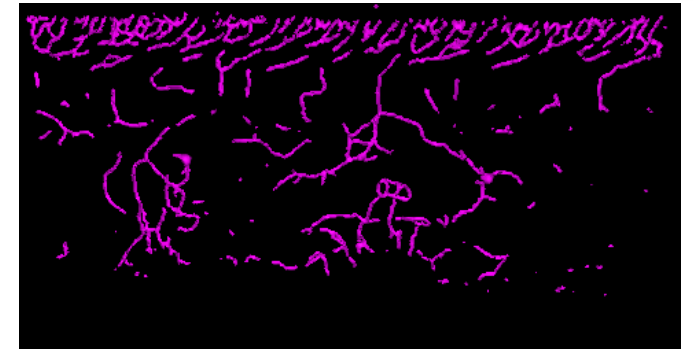
qimtools python package
<https://qim.dk/>



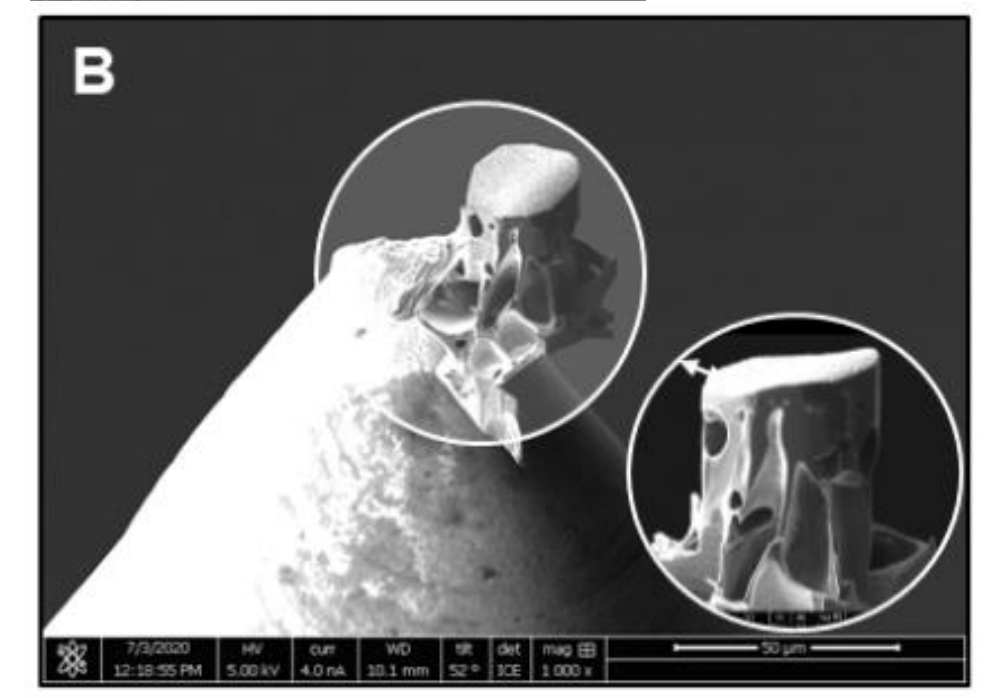
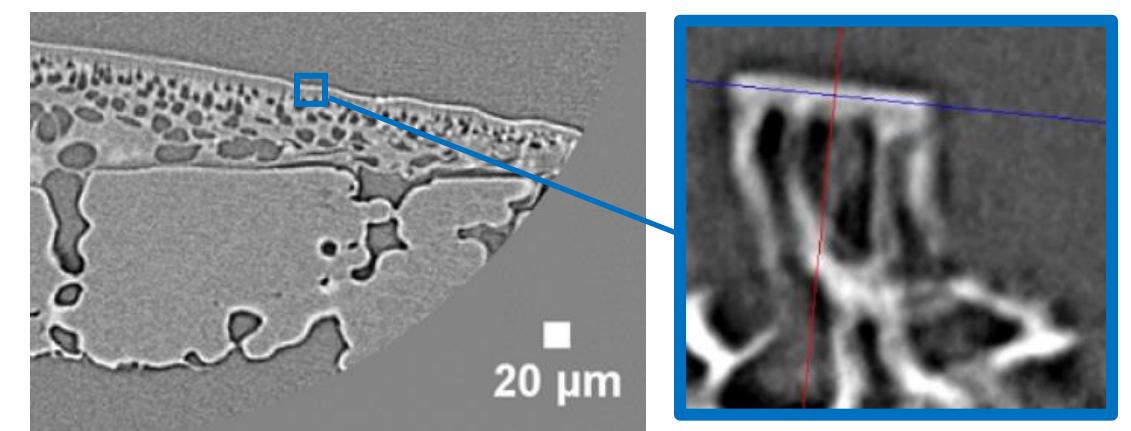
Phase retrieval carried out by Frans Matsson & Rajmund Mokso using an in-house developed Python script.



Image processing and analysis work is ongoing.

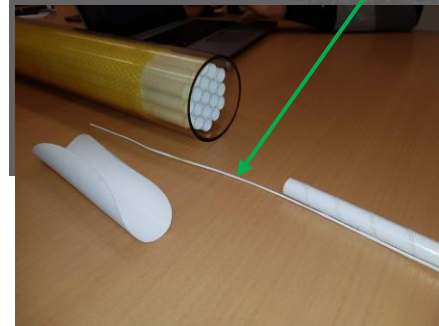
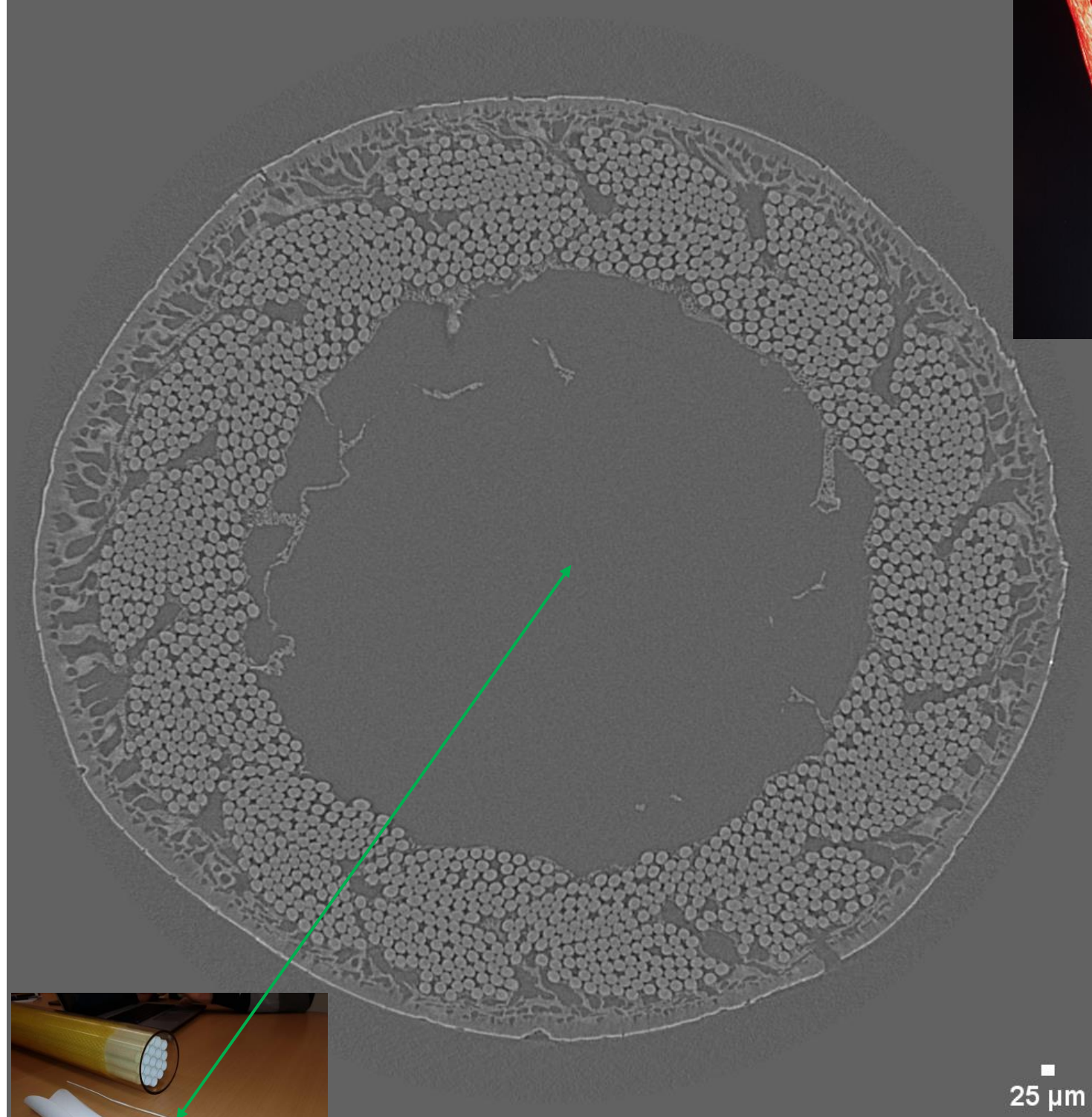
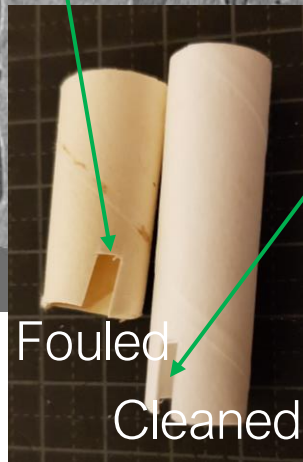
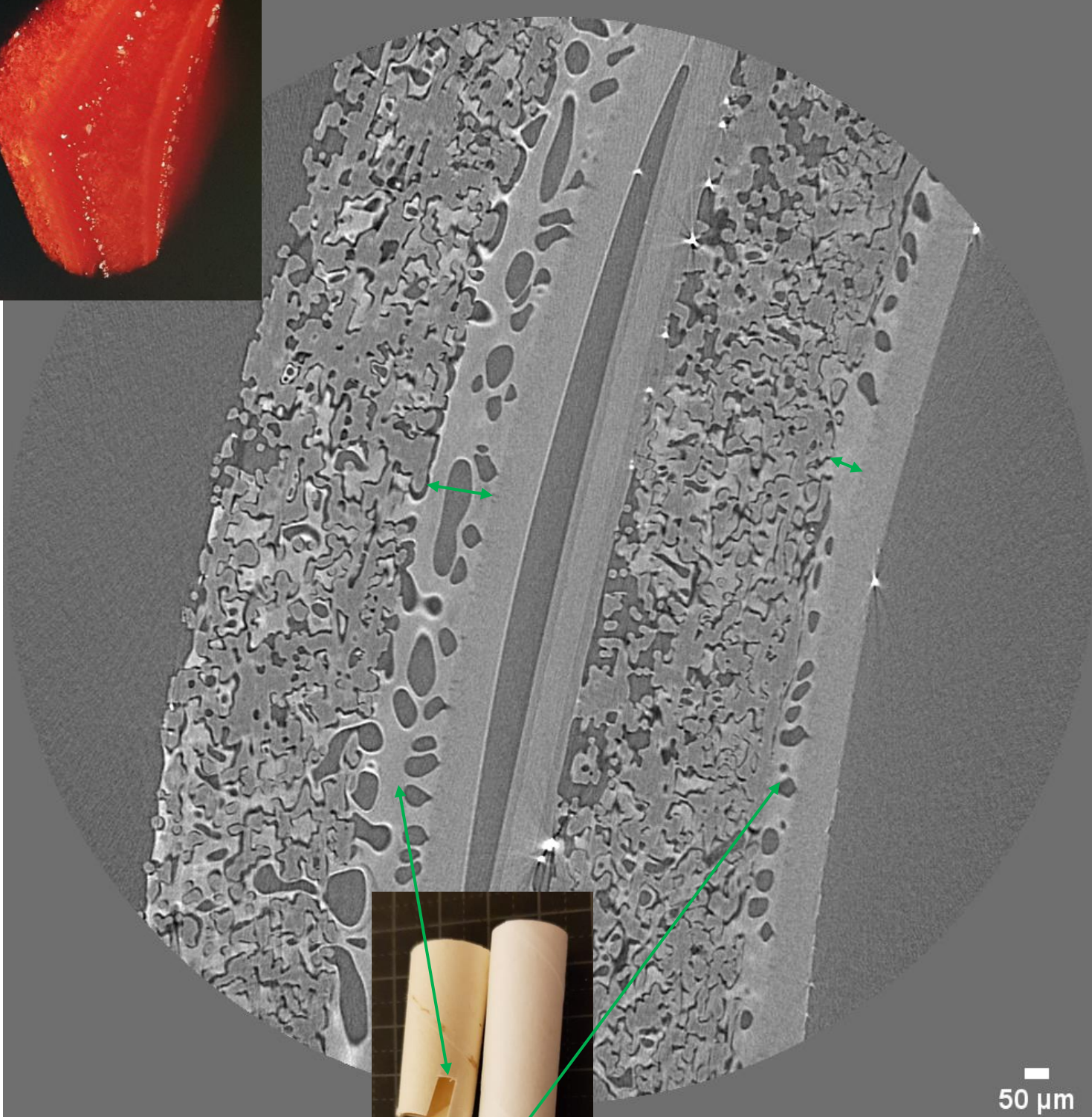


mCT scans of polysulfone-based membranes



Sample preparation by FIB:ing (Focused Ion Beam) carried out by Torben Nilsson Pingel, RISE/Chalmers.

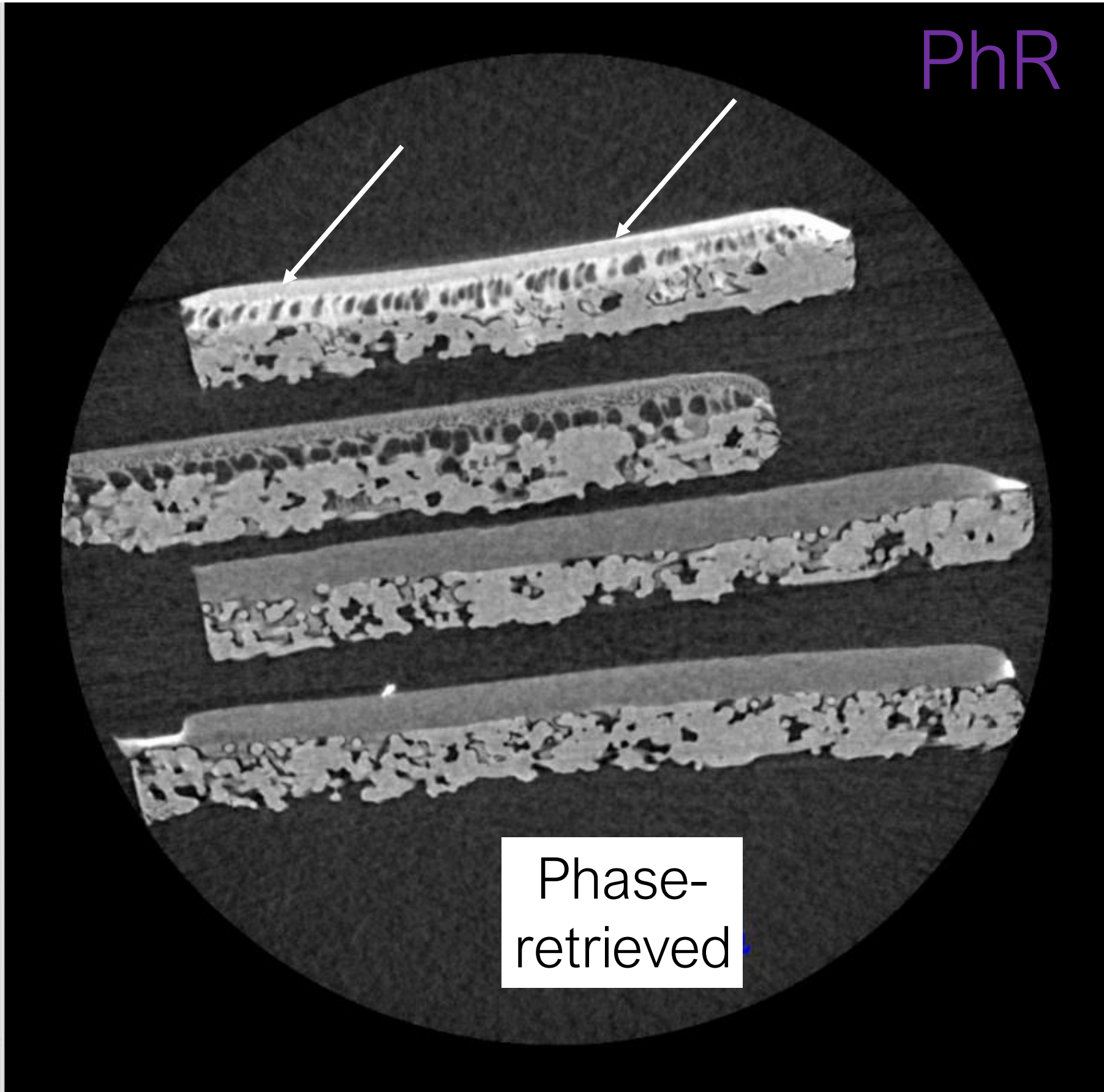
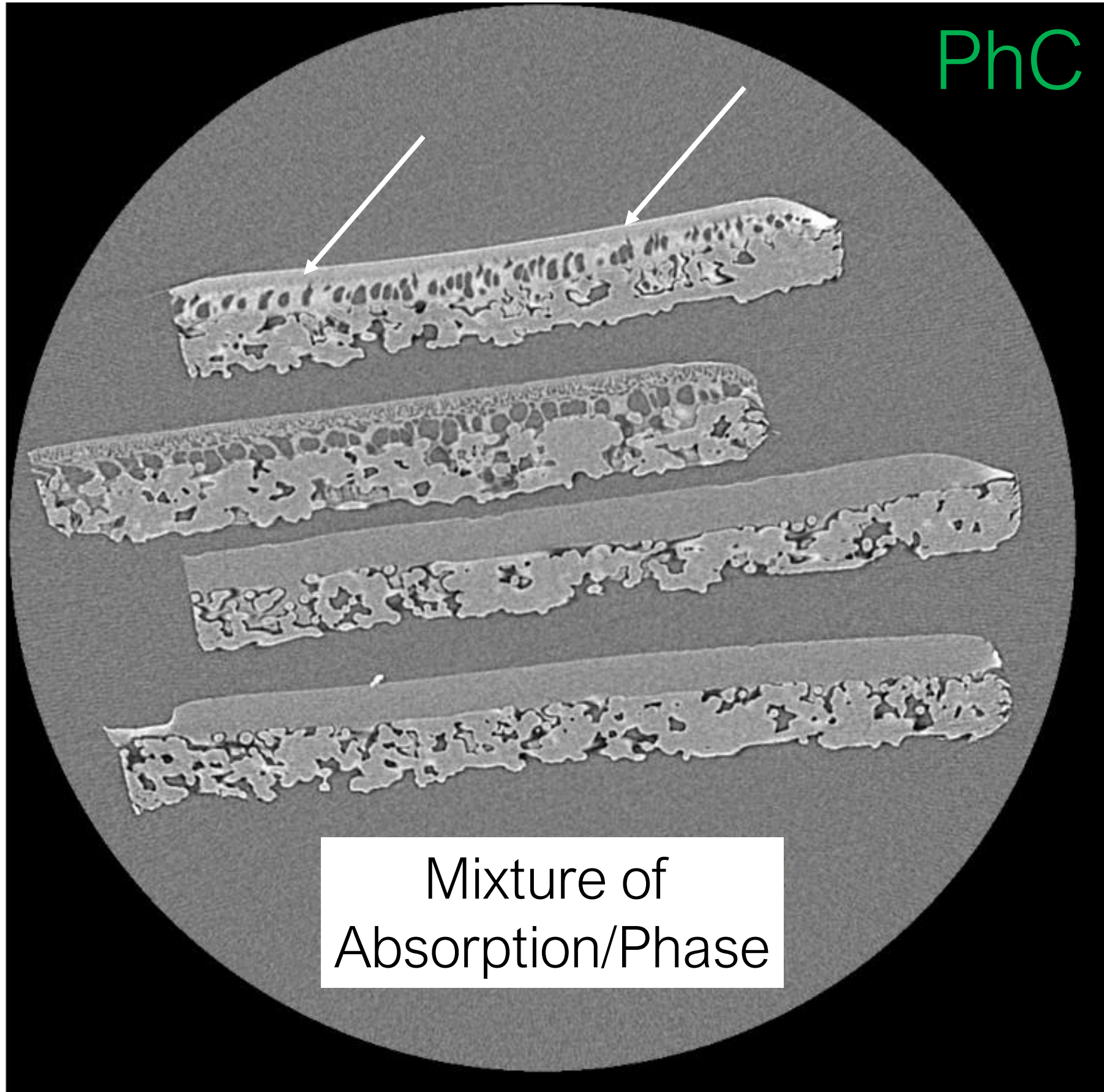
X-ray tomography of various membranes



Non-phase retrieved vs phase retrieved

PhC

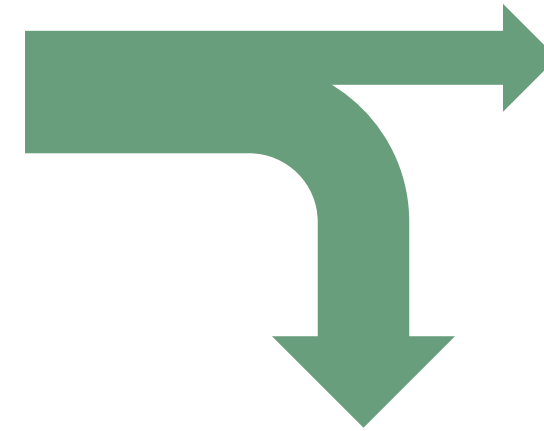
PhR



Rapeseed processing



3 kg
Seeds



2 kg
Presscake
≈30 % Protein



1 kg
Oil



Great amino
acid composition

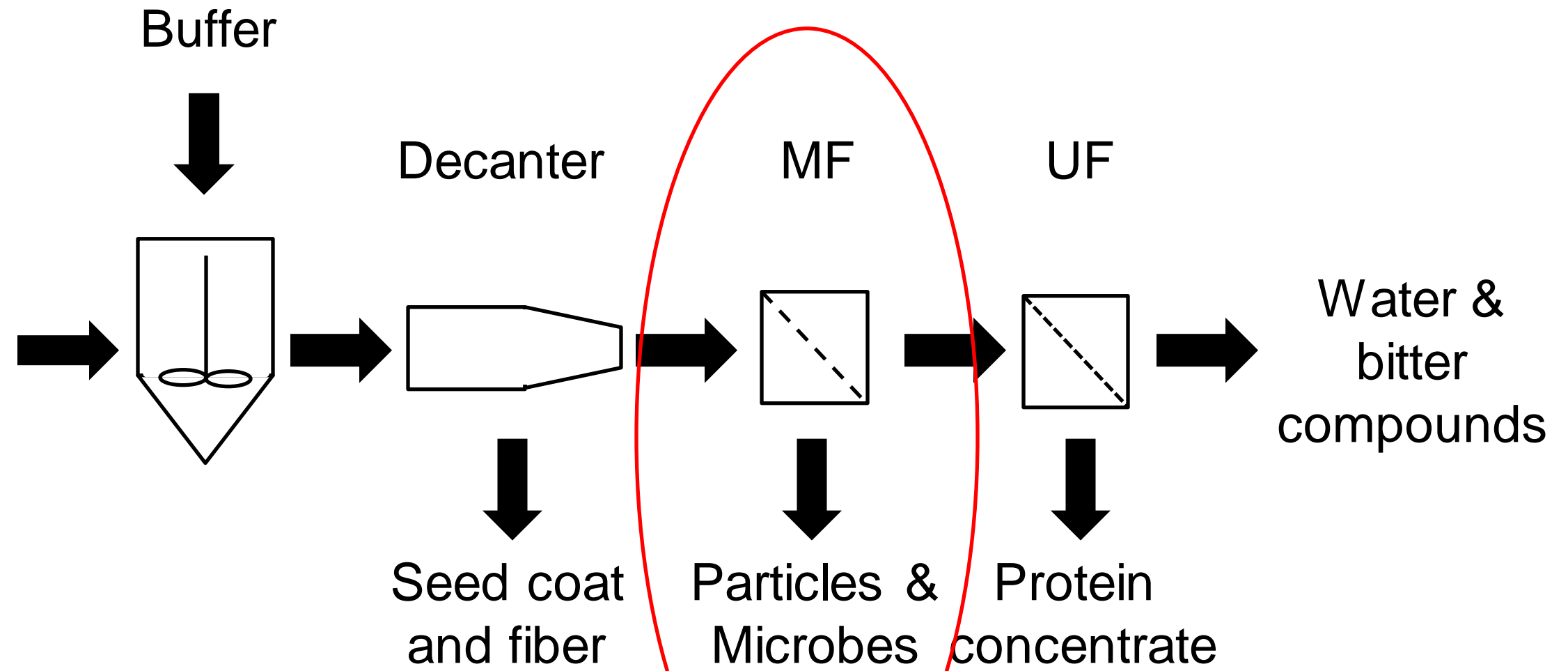
But bitter flavour

- World's second largest cultivated oilseed
- Rapeseed protein is comparable with soy protein in nutritional value
- Approved rapeseed protein isolated for human consumption in 2013 by European Food and Safety Authority

Rapeseed processing



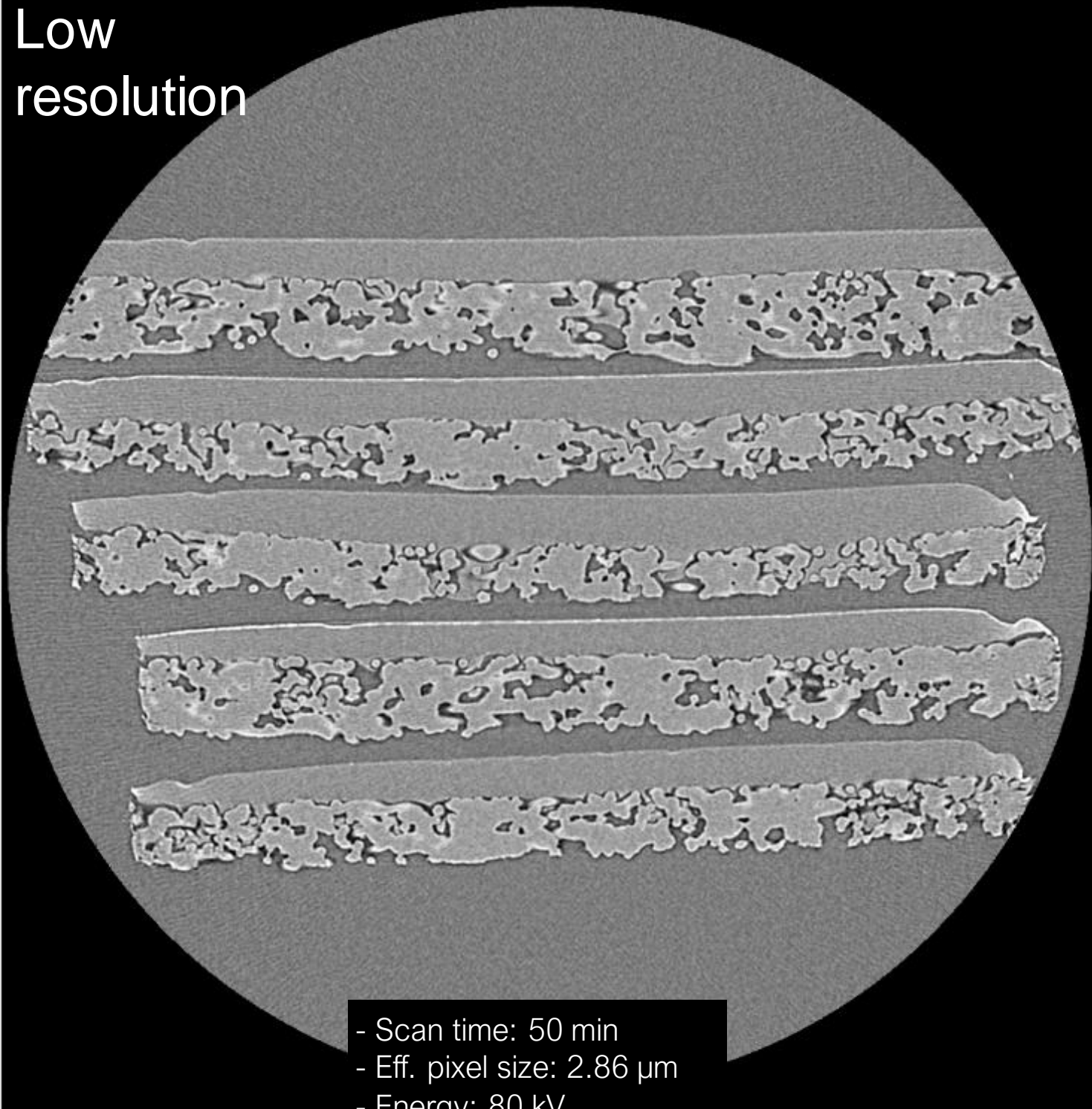
Presscake



Step studied

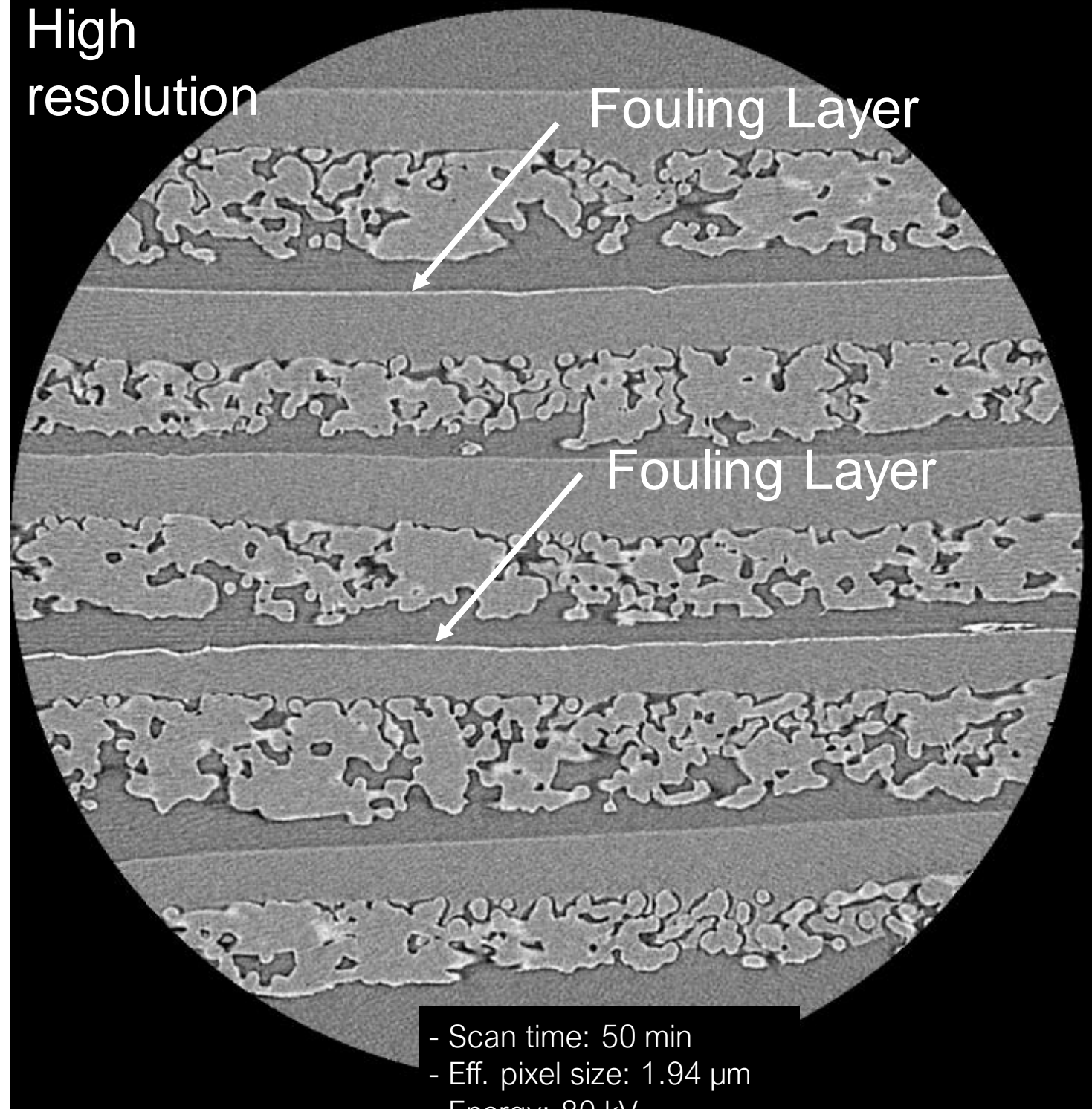
Lab μ CT scans of fouled and cleaned MF membranes

Low resolution



- Scan time: 50 min
- Eff. pixel size: 2.86 μ m
- Energy: 80 kV
- No. Proj: 1001
- Exp. Time: 1 s

High resolution



- Scan time: 50 min
- Eff. pixel size: 1.94 μ m
- Energy: 80 kV
- No. Proj: 1001
- Exp. Time: 1 s

MFG2 Pristine

MFG2 Fouled

MFG2 Cleaned

MFG1 Fouled

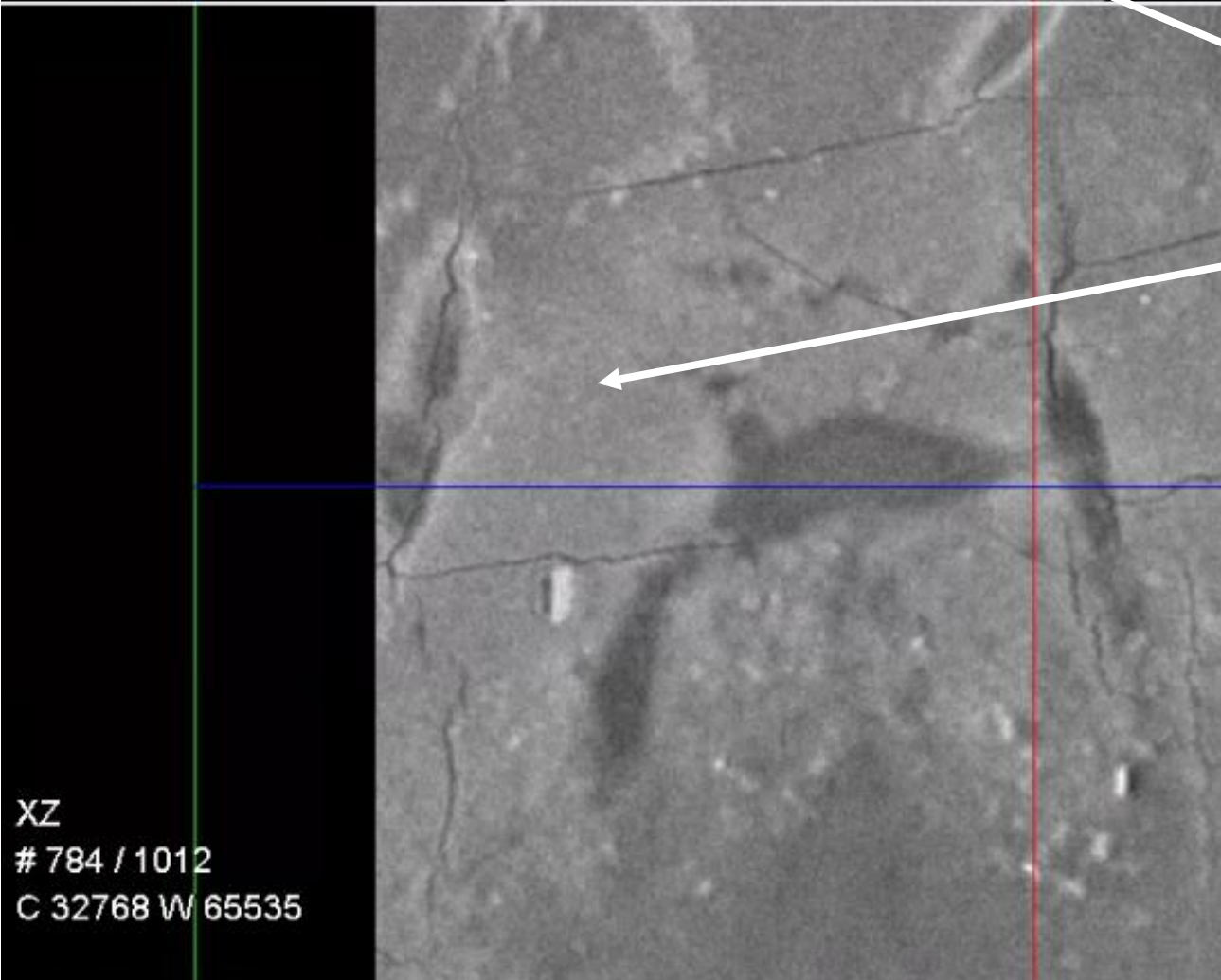
MFG1 Cleaned

MFG2 Fouled

Acquired: 9th of June
2021
- Scan time: 4 hours
- Eff. pixel size: 0.45 μm
- Energy: 80 kV
- No. Proj: 1001
- Exp. Time: 12 s

XY
498 / 983
C 32768 W 65535

YZ
617 / 864
C 32768 W 65535



Fouling
Layer

Non-woven
support

Take home messages

- X-ray tomographic imaging provides novel insight into the impact of membrane fouling and cleaning on the membrane structure
- High resolution available at synchrotron facilities is needed for detailed analysis
- The technology requires a wide network of experts and collaboration



Thank you for listening!

**RI
SE**

Dr. Shun Yu
Dr. Torben Nilsson Pingel



LUND
UNIVERSITY

Mr. Gregor Rudolph
Dr. Johan Thuvander
Dr. Emanuel Larsson
Assoc. Prof Stephen Hall
Assoc. Prof. Pablo Villanueva-Perez
Prof. Frank Lipnizki

VINNOVA
Sweden's Innovation Agency

Call: Increasing capacity and skills of PhD students regarding industrially relevant neutron and synchrotron-based analytical methods – 2019

Project: Measurements of membrane fouling in lignocellulosic biorefineries by ptycho-tomography, 2019-03613

Contact details:
gregor.rudolph@chemeng.lth.se
emanuel.larsson@solid.lu.se

PhD defence tomorrow on membrane fouling and cleaning

**Investigations on Membrane Fouling
and Cleaning in Ultrafiltration Processes
in Lignocellulosic Biorefineries**

GREGOR RUDOLPH | CHEMICAL ENGINEERING | LUND UNIVERSITY

When: 11 June, 9.15 am

Where: Livestreamed on YouTube
tinyurl.com/MembraneFouling

